

Prodrug Linker

Field

- 5 The present invention is directed to polymeric prodrugs having temporary linkages to amino groups of biologically active entities such as peptides, proteins, natural products or synthetic chemical compounds.

Background

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Typically, polymers are either used in a non-covalent fashion, with the drug compound physicochemically formulated into a solvent-polymer mixture, or by permanent covalent attachment of a polymer reagent to one of the drug's functional groups.

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- Non-covalent drug encapsulation has been applied to depot formulations for long-acting release profiles. Typically, drug is mixed with polymer material and processed in such fashion, that the drug becomes distributed throughout the bulk polymer material. Such polymer-protein aggregates may be shaped as microparticles which are administered as an injectable suspension or they are formulated as gels which are administered in a single bolus injection. Drug release occurs when the polymer swells or degradation of the polymer allows for diffusion of the drug to the exterior. Such degradation processes may be autohydrolytic or enzyme-catalyzed. An example for a marketed drug based on bolus administration of a drug-polymer gel is Lupron Depot.
- 20
- 25 An example for a marketed drug based on suspended microparticles is Nutropin Depot.

- A disadvantage of the non-covalent approach is that in order to prevent uncontrolled, burst-type release of the drug, encapsulation has to be highly efficient by creating a sterically highly crowded environment. Restraining the diffusion of an unbound, water soluble drug molecule requires strong van der Waals contacts, frequently mediated through hydrophobic moieties. Many conformationally sensitive therapeutics such as proteins or peptides are rendered dysfunctional during the encapsulation process and/or during subsequent storage. In addition, such amino-
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containing drug compounds readily undergo side reactions with polymer degradation products (D.H. Lee et al., J. Contr. Rel., 2003, 92, 291-299). Furthermore, dependence of the release mechanism upon biodegradation may cause interpatient variability.

- 5 Alternatively, drugs may be conjugated to polymers through permanent covalent bonds. This approach is applied to various classes of molecules, from so-called small molecules, through natural products up to larger proteins.

Many small molecule medicinal agents, like alkaloids and anti-tumor agents, show
10 low solubility in aqueous fluids. One way to solubilize these small molecule compounds is to conjugate them to hydrophilic polymers. A variety of water-soluble polymers, such as human serum albumin, dextran, lectins, poly(ethylene glycol) (PEG), poly(styrene-co-maleic anhydride), poly(N-hydroxypropylmethacrylamide), poly(divinyl ether-co-maleic anhydride), hyaluronic acid have been described for this
15 purpose (R. Duncan, Nature Rev. Drug Disc., 2003, 2, 347-360).

A major challenge in cancer therapy is to selectively target cytotoxic agents to tumor cells. A promising method to accumulate small molecule anticancer agents in tumor tissue and decrease undesirable side effects of these agents is the attachment of the
20 cytotoxin to a macromolecular carrier. The passive targeting of polymeric drug conjugates to tumors is based on the so-called enhanced permeability and retention effect (EPR) as described by Matsumura, Y. and Maeda, H., in Cancer Res., 1986, vol 6, pp 6387-6392. As a result, several polymer-drug conjugates have entered clinical trial as anticancer agents.

25 Covalent modification of biological molecules with poly(ethylene glycol) has been extensively studied since the late 1970s. So-called PEGylated proteins have shown improved therapeutic efficacy by increasing solubility, reducing immunogenicity, and increasing circulation half-life in vivo due to reduced renal clearance and proteolysis
30 by enzymes (see, for example, Caliceti P., Veronese F.M., Adv. Drug Deliv. Rev. 2003, 55, 1261-1277).

However, many medicinal agents such as INFalfa2, saquinavir or somatostatin are inactive or show decreased biological activity when a polymer is covalently

conjugated to the drug molecule (T. Peleg-Shulman et al., J. Med. Chem., 2004, 47, 4897-4904).

In order to avoid shortcomings imposed by either non-covalent polymer mixtures or permanent covalent attachment, it may be preferable to employ a prodrug approach for chemical conjugation of drug to polymer carrier. In such polymeric prodrugs, the biologically active moieties are typically linked to the polymeric carrier moiety by a temporary bond formed between the carrier moiety and a hydroxy, amino or carboxy group of the drug molecule (such as is shown in Fig. 1).

Prodrugs are therapeutic agents that are almost inactive per se but are predictably transformed into active metabolites (see B. Testa, J.M: Mayer in Hydrolysis in Drug and Prodrug Metabolism, Wiley-VCH, 2003, page 4). The carrier prodrug approach may be applied in such a fashion that the medicinal agent is released in vivo from the polymer in order to regain its biological activity. The reduced biological activity of the prodrug as compared to the released drug is of advantage if a slow or controlled release of the drug is desired. In this case, a relatively large amount of prodrug may be administered without concomitant side effects and the risk of overdosing. Release of the drug occurs over time, thereby reducing the necessity of repeated and frequent administration of the drug.

Prodrug activation may occur by enzymatic or non-enzymatic cleavage of the temporary bond between the carrier and the drug molecule, or a sequential combination of both, i.e. an enzymatic step followed by a non-enzymatic rearrangement. In an enzyme-free in vitro environment such as an aqueous buffer solution, a temporary bond such as an ester or amide may undergo hydrolysis, but the corresponding rate of hydrolysis may be much too slow and not therapeutically useful. In an in vivo environment, esterases or amidases are typically present and may cause significant catalytic acceleration of the kinetics of hydrolysis from twofold up to several orders of magnitude (see, for example, R.B. Greenwald et al. J.Med.Chem. 1999, 42 (18), 3857-3867).

Definitions based on IUPAC

(as given under <http://www.chem.qmul.ac.uk/iupac/medchem/> (accessed on 8 March 2004))

5 Prodrug

A prodrug is any compound that undergoes biotransformation before exhibiting its pharmacological effects. Prodrugs can thus be viewed as drugs containing specialized non-toxic protective groups used in a transient manner to alter or to eliminate undesirable properties in the parent molecule.

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Carrier-linked prodrug (Carrier prodrug)

A carrier-linked prodrug is a prodrug that contains a temporary linkage of a given active substance with a transient carrier group that produces improved physicochemical or pharmacokinetic properties and that can be easily removed in vivo, usually by a hydrolytic cleavage. This is shown graphically in Fig. 1.

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Cascade prodrug

A cascade prodrug is a carrier prodrug for which the cleavage of the carrier group becomes effective only after unmasking an activating group.

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Polymeric cascade prodrug

A polymeric cascade prodrug is a carrier prodrug that contains a temporary linkage of a given active substance with a transient polymeric carrier group for which the cleavage of the carrier becomes effective only after unmasking an activating group.

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Bioprecursor prodrug

A bioprecursor prodrug is a prodrug that does not imply the linkage to a carrier group, but results from a molecular modification of the active principle itself. This modification generates a new compound, able to be transformed metabolically or chemically, the resulting compound being the active principle.

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Biotransformation

Biotransformation is the chemical conversion of substances by living organisms or enzyme preparations.

Prodrugs fall in two classes, bioprecursors and carrier-linked prodrugs. Bioprecursors do not contain a carrier group and are activated by the metabolic creation of a functional group. In carrier-linked prodrugs the active substance is linked to a carrier moiety by a temporary linkage. This invention is concerned with polymeric carrier-linked or macromolecular prodrugs, where the carrier itself is a macromolecule such as a carrier protein or polysaccharide or polyethylene glycol. Specifically, the invention relates to polymeric carrier-linked prodrugs for which this cleavage between polymer and drug proceeds in two steps according to a cascade mechanism.

Cleavage of a carrier prodrug generates a molecular entity (drug) of increased bioactivity and at least one side product, the carrier. This side product may be biologically inert (for instance PEG) or may have targeting properties (for instance antibodies). After cleavage, the bioactive entity will reveal at least one previously conjugated and thereby protected functional group, and the presence of this group typically contributes to the drug's bioactivity.

In order to implement a prodrug strategy, at least one certain functional group in the drug molecule is employed for attachment of the carrier polymer. Preferred functional groups are hydroxyl or amino groups. Consequently, both the attachment chemistry and hydrolysis conditions vary greatly between these two functionalities.

In a simple one-step mechanism, the prodrug's temporary linkage is characterized by an intrinsic lability or enzyme dependence. The susceptibility of this linkage to hydrolysis in an aqueous environment with or without enzyme catalysis controls the cleavage kinetics between polymeric carrier and drug. Numerous macromolecular prodrugs are described in the literature where the temporary linkage is a labile ester bond. In these cases, the functional group provided by the bioactive entity is either a hydroxyl group or a carboxylic acid (e.g. Y. Luo, MR Ziebell, GD Prestwich, "A Hyaluronic Acid – Taxol Antitumor Bioconjugate Targeted to Cancer Cells", Biomacromolecules 2000, 1, 208-218, J Cheng et al, Synthesis of Linear, beta-Cyclodextrin Based Polymers and Their Camptothecin Conjugates, Bioconjugate Chem. 2003, 14, 1007-1017, R. Bhatt et al, Synthesis and in Vivo Antitumor Activity of Poly(L-glutamic acid) Conjugates of 20(S)-Camptothecin, J. Med. Chem. 2003,

46, 190-193 ; R.B. Greenwald, A. Pendri, C.D. Conover, H. Zhao, Y.H. Choe, A. Martinez, K. Shum, S. Guan, J. Med. Chem., 1999, 42, 3657-3667; B. Testa, J.M: Mayer in Hydrolysis in Drug and Prodrug Metabolism, Wiley-VCH, 2003, Chapter 8).

5 Especially for therapeutic biomacromolecules but also for certain small molecule drugs, it may be desirable to link the macromolecular carrier to amino groups of the bioactive entity (i.e. N-terminus or lysine amino groups of proteins). This will be the case if masking the drug's bioactivity requires conjugation of a certain amino group of the bioactive entity, for instance an amino group located in an active center or a
10 region or epitope involved in receptor binding. Also, during preparation of the prodrug, amino groups may be more chemoselectively addressed and serve as a better handle for conjugating carrier and drug because of their greater nucleophilicity as compared to hydroxylic or phenolic groups. This is particularly true for proteins which may contain a great variety of different reactive functionalities, where non-
15 selective conjugation reactions lead to undesired product mixtures which require extensive characterization or purification and may decrease reaction yield and therapeutic efficiency of the product.

Amide bonds as well as aliphatic carbamates are much more stable towards hydrolysis
20 than ester bonds, and the rate of cleavage would be too slow for therapeutic utility in a carrier-linked prodrug. Therefore it is advantageous to add structural chemical components such as neighbouring groups in order to exert control over the cleavability of the prodrug amide bond. Such additional cleavage-controlling chemical structures that are not provided by the carrier entity nor by the drug are
25 called linker. Prodrug linkers can have a strong effect on the rate of hydrolysis of a given temporary bond. Variation of the chemical nature of these linkers allows to engineer the linker properties to a great extent.

For instance, prodrug linkers may be designed for enzyme-selectivity. Prerequisite for
30 enzymatic dependence is that the linker structure displays a structural motif that is recognized as a substrate by a corresponding endogenous enzyme (Fig. 2).

Enzyme-catalyzed acceleration of prodrug cleavage is a desirable feature for organ or cellular targeting applications. Targeted release of the bioactive entity is effected, if

an enzyme, that selectively cleaves the linkage, is specifically present in the organ or cell-type chosen for treatment.

A typical property of an enzyme-dependent temporary linkage is its stability with respect to hydrolysis. The temporary linkage itself will not undergo autohydrolysis at a rate that would release drug to such an extent that a therapeutic effect could be induced in a normal dosing regime. It is only in the presence of the enzyme, that the attack of the enzyme on the linkage causes a significant acceleration of cleavage and concomitant an enhancement of free drug concentration.

Several examples have been published for the prodrug activation of amine-containing biologically active moieties by specific enzymes for targeted release. In these cases, cleavage occurs in a one-step process which is catalyzed by the enzyme. G. Cavallaro et al., *Bioconjugate Chem.* 2001, 12, 143-151 describe the enzymatic release of an antitumoral agent by the protease plasmin. Cytarabin is coupled via the tripeptide sequence D-Val-Leu-Lys to the polymer alpha, beta-poly(N-hydroxyethyl)-DL-aspartamide (PHEA). Enzymatic release of cytarabin is effected by the protease plasmin which concentration is relatively high in various kinds of tumor mass.

Further examples for antitumoral polymeric prodrugs activated by specific enzymes like beta lactamase (R. Satchi-Fainaro et al., *Bioconjugate Chem.* 2003, 14, 797-804) and cysteine proteases like cathepsin B (R. Duncan et al. *J. Contr. Release* 2001, 74, 135-146) have been described. Wiwattanapatapee et al. (2003) outline a dendrimer prodrug for colonic delivery of 5-aminosalicylic acid. The drug molecule is conjugated by an azo bond to "generation 3" PAMAM dendrimer. 5-aminosalicylic acid is released in the colon by a bacterial enzyme called azo reductase (W. R. Wiwattanapatapee, L. Lomlim, K. Saramunee, *J. Controlled Release*, 2003, 88: 1-9).

A.J. Garman et al. (A.J. Garman, S.B. Kalindjan, *FEBS Lett.* 1987, 223 (2), 361-365 1987) use PEG5000-maleic anhydride for the reversible modification of amino groups in tissue-type plasminogen activator and urokinase. Regeneration of functional enzyme from PEG-uPA conjugate upon incubation at pH 7.4 buffer by cleavage of the maleamic acid linkage follows first order kinetics with a half-life of 6.1 h. The prodrug cleavage was not investigated in the presence of enzymes, and it can be

expected – as explained above - that proteases present in the in vivo environment will significantly contribute to the cleavage of the temporary amide linkage. A further disadvantage of this linkage is the lack of stability of the conjugate at lower pH values. This limits the applicability of the linker to active agents which are stable at basic pH values, as purification of the active agent polymer conjugate has to be performed under basic conditions to prevent premature prodrug cleavage.

Cascade mechanisms have proven particularly useful in the controlled release of drugs containing amino-group functionalities because linker cleavage characteristics can be optimized with greater flexibility than in simple one-step prodrugs.

Cascade cleavage is enabled by linker compounds that are composed of a structural combination of a masking group and an activating group. The masking group is attached to the activating group by means of a first temporary linkage such as an ester or a carbamate. The activating group is attached to an amino-group of the drug molecule through a second temporary linkage, for instance a carbamate. The stability, or susceptibility to hydrolysis of the second temporary linkage is dependent on the presence or absence of the masking group. In the presence of the masking group, the second temporary linkage is highly stable and unlikely to release drug with therapeutically useful kinetics. In the absence of the masking group, this linkage becomes highly labile, causing rapid cleavage and drug release.

Cleavage of the first temporary linkage is the rate-limiting step in the cascade mechanism. This first step may induce a molecular rearrangement of the activating group such as a 1,6-elimination. The rearrangement renders the second temporary linkage so much more labile that its cleavage is induced. Ideally, the cleavage rate of the first temporary linkage is identical to the desired release rate for the drug molecule in a given therapeutic scenario. Furthermore, it is desirable that cleavage of the second temporary linkage is instantaneous after its lability has been induced by cleavage of the first temporary bond.

A variety of examples exist for cascade carrier prodrugs where the masking group functionality is performed by the carrier polymer itself as shown diagrammatically in Fig. 3. In the systems discussed below, the masking group is not only part of the

carrier but has also been engineered for enzyme-dependence (Fig. 4). Only in the presence of a corresponding enzyme is the rate of cleavage of the first temporary linkage sufficiently accelerated for therapeutic use.

- 5 R.B. Greenwald, A. Pendri, C.D. Conover, H. Zhao, Y.H. Choe, A. Martinez, K. Shum, S. Guan, J. Med. Chem., 1999, 42, 3657-3667 & PCT Patent Application WO-A-99/30727 describe a methodology for synthesizing poly(ethylene glycol) prodrugs of amino-containing small molecule compounds based on 1,4- or 1,6-benzyl elimination. In this approach, poly(ethylene glycol) as the polymeric carrier is
10 attached to the benzyl group by means of a first temporary linkage such as an ester, carbonate, carbamate, or amide bond. The benzyl group serves as the activating group, and the PEG polymer also has the function of the masking group in this cascade cleavage mechanism. The amino group of the drug molecule is linked via a second temporary linkage, containing a carbamate group, to the benzyl moiety. The
15 release of PEG from the drug molecule is initiated by enzymatic cleavage of the first temporary linkage followed by a rapid 1,4- or 1,6-benzyl elimination, initiating cleavage of the second temporary linkage.

The same linker system is also used for releasable poly(ethylene glycol) conjugates of
20 proteins (S. Lee, R.B. Greenwald et al. Bioconj. Chem. 2001, 12 (2), 163-169). Lysozyme is used as model protein because it loses its activity when PEGylation takes place on the epsilon-amino group of lysine residues. Various amounts of PEG linker were conjugated to the protein. Regeneration of native protein from the PEG conjugates occurs by enzymatic cleavage in rat plasma or in non-physiological high
25 pH buffer.

Greenwald et al. published in 2000 a poly(ethylene glycol) drug delivery system of amino-containing prodrugs based on trimethyl lock lactonization (R.B. Greenwald et al. J. Med. Chem. 2000, 43(3), 457-487; PCT Patent Application No. WO-A-
30 02/089789). In this prodrug system, substituted o-hydroxyphenyl-dimethylpropionic acid is linked to PEG by an ester, carbonate, or carbamate group as a first temporary linkage and to amino groups of drug molecules by means of an amide bond as second temporary linkage. The rate-determining step in drug release is the enzymatic

cleavage of the first linkage. This step is followed by fast amide cleavage by lactonization, liberating a potentially toxic aromatic lactone side product.

Similar prodrug systems were described by F.M.H. DeGroot et al. (WO02083180 and
5 WO04043493A1) and D. Shabat et al. (WO04019993A1). WO02083180 discloses a prodrug system with elongated and multiple linkers based on 1,(4+2n) elimination. The masking moieties in these examples were specifically designed for enzymatic cleavage. This approach was extended to dendritic prodrug system where one enzymatic activating event triggered the release of more than one drug molecule
10 (WO04043493A1). WO04019993A1 discloses a similar prodrug system based on a self-immolative dendrimer releasing many drug moieties upon a single enzymatic activating event. These systems are characterized by the absence of a polymeric carrier. Instead, oligomerization of prodrug linker components provides for a high molecular weight of the prodrug, and prodrug cleavage generates linker residues and
15 free drug, but no polymeric entity is released.

The disadvantage in the abovementioned prodrug systems described by Greenwald, DeGroot and Shabat is the release of potentially toxic aromatic small molecule side products like quinone methides after cleavage of the temporary linkage. The
20 potentially toxic entities are released in a 1:1 stoichiometry with the drug and can assume high in vivo concentrations. This risk factor is even greater if self-immolative dendritic structures based on oligomers of the activating group are employed and more aromatic side products than drug molecules are released.

25 More recently, R.B. Greenwald et al. (Greenwald et al. J. Med.Chem. 2004, 47, 726-734) described a PEG prodrug system based on bis-(N-2-hydroxyethyl)glycin amide (bicin amide) linker. In this system two PEG molecules are linked to a bicin molecule coupled to an amino group of the drug molecule. The first two steps in prodrug activation is the enzymatic cleavage of both PEG molecules. Different linkages
30 between PEG and bicin are described resulting in different prodrug activation kinetics. The main disadvantage of this system is the slow hydrolysis rate of bicin amide conjugated to the drug molecule ($t_{1/2} = 3$ h in phosphate buffer) which results in the release of a bicin-modified prodrug intermediate that may show different

pharmacokinetic and pharmacodynamic properties as compared to the parent drug molecule.

5 Cascade prodrugs with masking groups that are part of the carrier polymer are limited in the control of drug release kinetics. As masking group cleavage is the rate-limiting step in the cascade mechanism, its molecular structure governs the kinetics. If the carrier polymer is identical to the masking group, the structural flexibility is restricted to the polymers' features. Alternatively, if the polymer requires structural modification in order to match the requirements for controlled cleavage, synthesis of
10 corresponding structures may become more difficult. Also, the incorporation of masking group features into a polymer may change its safety profile.

Therefore it is preferred to structurally separate the masking group and the carrier. This may be achieved by employing a permanent bond between polymer carrier and
15 activating group. This stable bond does not participate in the cascade cleavage mechanism. If the carrier is not serving as a masking group and the activating group is coupled to the carrier by means of a stable bond, release of potentially toxic side products such as the activating group is avoided. The stable attachment of activating group and polymer also suppresses the release of drug-linker intermediates with
20 undefined pharmacology.

Systems have been developed for targeted delivery of therapeutic agents by rendering the masking group enzyme-dependent. Only in the presence of a corresponding enzyme is the rate of cleavage of the first temporary linkage connecting the masking
25 group with the activating group sufficiently accelerated for therapeutic use.

Antczak et al. (Bioorg Med Chem 9 (2001) 2843-48) describe a reagent which forms the basis for a macromolecular cascade prodrug system for amine-containing drug molecules. In this approach an antibody serves as carrier, a stable bond connects the
30 antibody to an activating moiety, carrying an enzymatically cleavable masking group. Upon enzymatic removal of the ester-linked masking group, a second temporary bond cleaves and releases the drug compound, as shown in Fig. 6.

D. Shabat et al. (Chem. Eur. J. 2004, 10, 2626-2634) describe a polymeric prodrug system based on a mandelic acid activating moiety. In this system the masking group is linked to the activating moiety by a carbamate bond. The activating moiety is conjugated permanently to a polyacrylamide polymer via an amide bond. After enzymatic activation of the masking group by a catalytic antibody, the masking group is cleaved by cyclization and the drug is released. The activating moiety is still connected to the polyacrylamide polymer after drug release.

M.-R. Lee et al. describe (Angew. Chem. 2004, 116, 1707-1710) a similar prodrug system based on mandelic acid activating moiety and an enzymatically cleavable ester-linked masking group.

In all of these described prodrug-polymer systems the masking group is specifically designed to be substrate to an enzyme, and masking group cleavage will almost entirely depend upon enzymatic catalysis with the disadvantages of interpatient variability, injection site variability and poor in vitro-in vivo correlation.

A major drawback of predominantly enzymatic cleavage is interpatient variability. Enzyme levels may differ significantly between individuals resulting in biological variation of prodrug activation by enzymatic cleavage. Enzyme levels may also vary depending on the site of administration, for instance it is known that in the case of subcutaneous injection, certain areas of the body yield more predictable therapeutic effects than others. To reduce this unpredictable effect, non-enzymatic cleavage or intramolecular catalysis is of particular interest (see, for example, B. Testa, J.M. Mayer in Hydrolysis in Drug and Prodrug Metabolism, Wiley-VCH, 2003, page 5).

Furthermore, it is difficult to establish an in vivo-in vitro correlation of the pharmacokinetic properties for such enzyme-dependent carrier-linked prodrugs. In the absence of a sound in vivo-in vitro correlation the optimization a release profile becomes a cumbersome task.

Also, the need for enzyme selectivity imposes a severe limitation on the structural features that can be used in the prodrug linker. This restriction greatly hinders the

development of a sound structure-activity relationship and consequently the optimization of linker cleavage kinetics.

For these reasons, there is a need to provide novel linker and/or carrier technologies
5 for forming polymeric prodrugs of amine containing active agents in order to overcome the limitations of the described polymeric prodrugs.

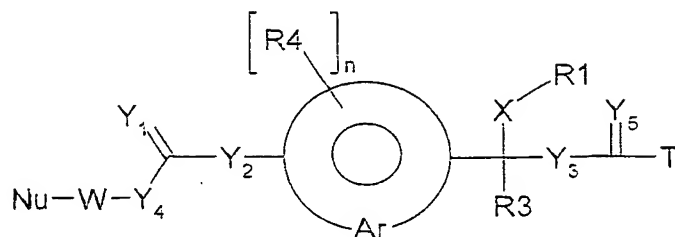
Detailed description of the invention

The present invention addresses the disadvantages described above. The invention provides for polymeric cascade prodrugs characterized by a masking group containing
5 a nucleophile and being distinct from the carrier.

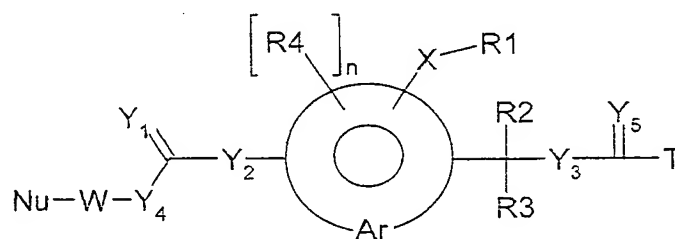
The nucleophile is in a suitable distance to a first temporary linkage with an aromatic activating group capable of undergoing a 1,(4+2p) elimination reaction (with $p = 0, 1, 2, 3, 4, \dots$) after cleavage of the first temporary linkage. The invention is further
10 characterized by the activating group being connected to the amino group of a drug molecule through a second temporary bond which is cleaved as a consequence of the 1,(4+2p) elimination. An additional structural feature is the attachment of a polymeric carrier to the activating group by means of a permanent bond.

15 The masking groups according to the present invention contain at least one nucleophile Nu. This nucleophile, which can be for example a primary, secondary or tertiary amino group can assist the cleavage of the masking group from the activating moiety by intramolecular catalysis or cyclization.

The invention provides for polymeric cascade prodrugs and corresponding polymeric cascade prodrug linker reagents of Formula Ia or Ib.



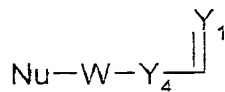
Formula Ia



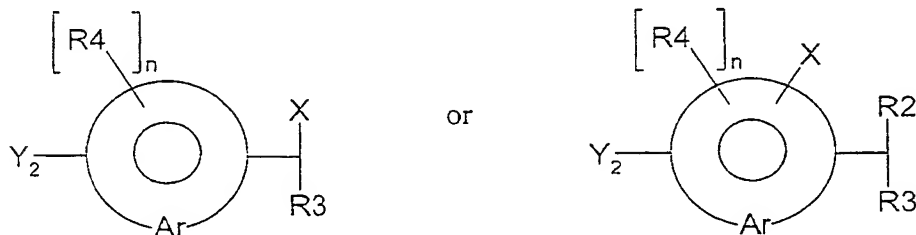
Formula Ib

wherein Y_1 to Y_5 , R_1 to R_4 , T , X , W , Nu and Ar are defined below:

Native drug release is effected by a two step mechanism. The first step is the rate-determining cleavage of the first temporary linkage between the masking group



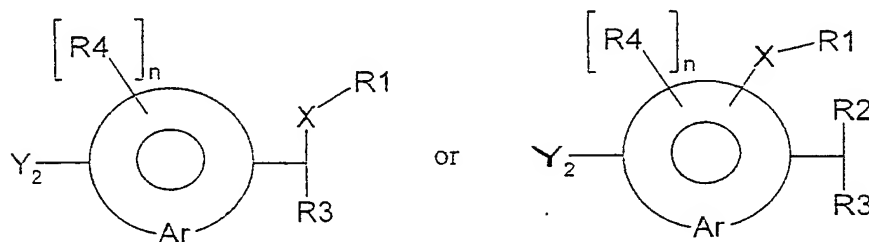
and the activating moiety



from the polymeric prodrug in vivo.

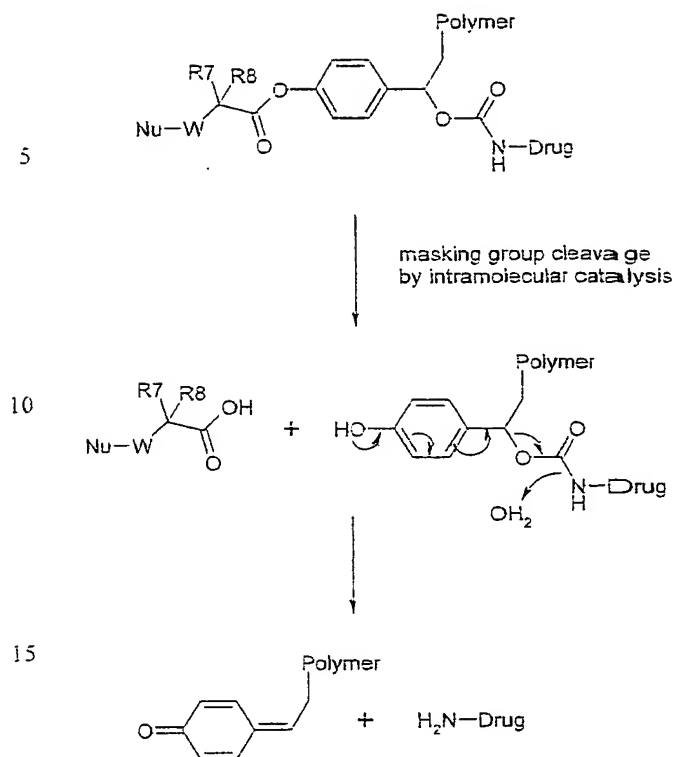
As described above, cleavage of the masking group may be mediated by an enzymatic or a non-enzymatic step, such as pH-dependent hydrolysis or intramolecular cyclization. In the preferred embodiment of the invention, cleavage is effected non-enzymatically by intramolecular cyclization or catalysis. The half-life of the cleavage kinetics in an aqueous buffer of pH 7.4 at 37 °C of the masking group according to the present invention is preferably between 1 hour and 6 months, more preferably between 1 day and 3 months, and most preferably between 1 day and 2 months.

The second and final step in release of the regenerated native drug is the fast, spontaneous and irreversible so-called 1,4- or 1,6 or 1,(4+2p) (in which p = 2, 3, 4 or higher) elimination of the



moiety of the remaining polymeric prodrug of formula Ia or formula Ib, respectively.

This mechanism of native drug release from a polymeric prodrug triggered by hydrolytic cleavage of the masking group followed by a 1,6-elimination step of the activating group is exemplified by a polymeric prodrug according to the present invention.



20 **Definition of Y_1 to Y_5 , R_1 to R_4 , T , X , W , Nu and Ar in formula Ia or Ib**

T is D or A

In the case where the inventive structure is a polymeric cascade prodrug linker reagent, T is A , and A is a leaving group. Non-limiting examples of suitable leaving groups A include but are not limited to chloride, bromide, fluoride, nitrophenoxy, imidazolyl, N -hydroxysuccinimidyl, N -hydroxybenzotriazolyl, N -hydroxyazobenzotriazolyl, pentafluorophenoxy, N -hydroxysulfosuccinimidyl, or any other leaving group known by those skilled in the art.

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In the case where the inventive structure is a polymeric cascade prodrug, T is D , and D is a residue of an amine-containing biologically active material including but not limited to small molecule bioactive agents or biopolymers like proteins, polypeptides and oligonucleotides (RNA, DNA), peptide nucleic acids (PNA),

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Note that in this description reference is often made to prodrugs. A true prodrug is found when T is the residue of the amine-containing biologically active material or moiety. If T is a leaving group A then the formula represents a polymeric cascade prodrug linker reagent. For simplicity these will be referred to prodrugs in this description. It will be understood from the context whether a true prodrug or a reagent as a precursor is meant.

Suitable organic small molecule bioactive moieties include, without limitation, moieties such as central nervous system-active agents, anti-infective, anti-neoplastic, antibacterial, anti-fungal, analgesic, contraceptive, anti-inflammatory, steroidal, vasodilating, vasoconstricting, and cardiovascular agents with at least one primary or secondary amino group. Non-exclusive examples of such compounds are daunorubicin, doxorubicin, idarubicin, mitoxantron, amino glutethimide, amantadine, diaphenylsulfon, ethambutol, sulfadiazin, sulfamerazin, sulfamethoxazol, sulfalen, cinafloxacin, moxifloxacin, ciprofloxacin, enoxacin, norfloxacin, neomycin B, spretnomycin, kanamycin A, meropenem, dopamin, dobutamin, lisinopril, serotonin, carbutamid, acivicin, etc.

Suitable proteins and polypeptides having at least one free amino group include but are not limited to ACTH, adenosine deaminase, agalsidase, albumin, alfa-1 antitrypsin (AAT), alfa-1 proteinase inhibitor (API), alteplase, anistreplase, anrod serine protease, antibodies (monoclonal or polyclonal, and fragments or fusions), antithrombin III, antitrypsins, aprotinin, asparaginases, biphalin, bone-morphogenic proteins, calcitonin (salmon), collagenase, DNase, endorphins, enfuvirtide, enkephalins, erythropoietins, factor VIIa, factor VIII, factor VIIIa, factor IX, fibrinolysin, fusion proteins, follicle-stimulating hormones, granulocyte colony stimulating factor (G-CSF), galactosidase, glucagon, glucagon-like peptides like GLP-1, glucocerebrosidase, granulocyte macrophage colony stimulating factor (GM-CSF), phospholipase-activating protein (PLAP), gonadotropin chorionic (hCG), hemoglobins, hepatitis B vaccines, hirudin, hyaluronidases, iduronidase, immune globulins, influenza vaccines, interleukins (1 alfa, 1 beta, 2, 3, 4, 6, 10, 11, 12), IL-1 receptor antagonist (rhIL-1ra), insulins, interferons (alfa 2a, alfa 2b, alfa 2c, beta 1a, beta 1b, gamma 1a, gamma 1b), keratinocyte growth factor (KGF), transforming growth factors, lactase, leuprolide, levothyroxine, luteinizing hormone, lyme vaccine,

natriuretic peptide, pancrelipase, papain, parathyroid hormone, PDGF, pepsin, platelet activating factor acetylhydrolase (PAF-AH), prolactin, protein C, octreotide, secretin, sermorelin, superoxide dismutase (SOD), somatropins (growth hormone), somatostatin, streptokinase, sucrase, tetanus toxin fragment, tilactase, thrombins, thymosin, thyroid stimulating hormone, thyrotropin, tumor necrosis factor (TNF), TNF receptor-IgG Fc, tissue plasminogen activator (tPA), TSH, urate oxidase, urokinase, vaccines, plant proteins such as lectins and ricins.

Also included herein is any synthetic polypeptide or any portion of a polypeptide with in vivo bioactivity. Furthermore, proteins prepared by recombinant DNA methodologies including mutant versions of aforementioned proteins, antibody fragments, single chain binding proteins, catalytic antibodies and fusion proteins are included.

Preferred proteins are antibodies, calcitonin, G-CSF, GM-CSF, erythropoietins, hemoglobins, interleukins, insulins, interferons, SOD, somatropin, TNF, TNF-receptor-IgG Fc, and GLP-1.

X is a spacer moiety such as R5-Y6

Y₁, Y₂ can each be either O, S, or NR6, independently of each other.

Y₃, Y₅ can each be either O or S, independently of each other.

Y₄ is O, NR6, or -C(R7)(R8)-

Y₆ is O, S, NR6, succinimide, maleimide, unsaturated carbon-carbon bonds or any

heteroatom containing a free electron pair, or is not present.

R2 and R3 are selected independently from hydrogen, substituted or non-substituted linear, branched or cyclical alkyl or heteroalkyl, aryls, substituted aryls, substituted or non-substituted heteroaryl, cyano, nitro, halogen, carboxy, carboxyalkyl, alkylcarbonyl, carboxamidoalkyl, etc.

The term "heteroalkyl" in the context of the present invention denotes (linear, cyclical or branched) alkyl chains where the alkyl chains contain or are substituted with at any position one or more heteroatoms, selected independently from O, S, N, P, Si, Cl, F, Br, I, etc. or groups, selected independently from carboxamide, carboxylic ester,

phosphonate ester, phosphate ester, double or triple bonds, carbamate, urea, thiourea, thiocarbamate, oxime, cyano, carboxyl, carbonyl, etc.

Each R₄ substitution on Ar may be the same or different and is selected independently
5 from hydrogen, substituted or non-substituted linear, branched or cyclical alkyl or
heteroalkyl, aryl, substituted aryl, substituted or non-substituted heteroaryl,
substituted or non-substituted linear, branched, or cyclical alkoxy, substituted or non-
substituted linear, branched, or cyclical heteroalkyloxy, aryloxy, heteroaryloxy,
cyano, halogen, etc.

10

R₄ is selected preferably from small substituents such as hydrogen, methyl, ethyl,
ethoxy, methoxy, and other C₁ to C₆ linear, cyclical or branched alkyls and
heteroalkyls.

15

n is zero or a positive integer.

R₇ and R₈ are selected independently from hydrogen, substituted or non-substituted
linear, branched or cyclical alkyl or heteroalkyl, aryls, substituted aryls, substituted or
non-substituted heteroaryl, cyano, halogen, etc.

20

R₅ is selected from substituted or non-substituted linear, branched or cyclical alkyl or
heteroalkyl, aryls, substituted aryls, substituted or non-substituted heteroaryl, etc.

25

R₆ is selected from hydrogen, substituted or non-substituted linear, branched or
cyclical alkyl or heteroalkyl, aryls, substituted aryls, substituted or non-substituted
heteroaryl, etc.

R₁ is a polymer.

30

Non-limiting examples for suitable polymers are polyalkyloxy-based polymers like
poly(propylene glycol) or poly(ethylene glycol), dextran, chitosan, hyaluronic acid
and derivatives, alginate, xylan, mannan, carrageenan, agarose, cellulose, starch,
hydroxyethyl starch (HES) and other carbohydrate-based polymers, poly(vinyl
alcohols), poly(oxazolines), poly(anhydrides), poly(ortho esters), poly(carbonates),
poly(urethanes), poly(acrylic acids), poly(acrylamides) such as

poly(hydroxypropylmethacrylamide) (HMPA), poly(acrylates), poly(methacrylates) like poly(hydroxyethylmethacrylate), poly(organophosphazenes), poly(siloxanes), poly(vinylpyrrolidone), poly(cyanoacrylates), poly(esters) such as poly(lactic acid) or poly(glycolic acids), poly(iminocarbonates), poly(amino acids) such as poly(glutamic acid), collagen, gelatin, copolymers, grafted copolymers, cross-linked polymers, hydrogels, and block copolymers from the above listed polymers.

Hydrogels may be defined as three-dimensional, hydrophilic or amphiphilic polymeric networks imbibing large quantities of water. The networks are composed of homopolymers or copolymers, are insoluble due to the presence of covalent chemical or physical (ionic, hydrophobic interactions, entanglements) crosslinks. The crosslinks provide the network structure and physical integrity. Hydrogels exhibit a thermodynamic compatibility with water which allows them to swell in aqueous media. (see.: N.A. Peppas, P. Bures, W. Leobandung, H. Ichikawa, Hydrogels in pharmaceutical formulations, Eur. J. Pharm. Biopharm. 2000, 50, 27-46). The chains of the network are connected in such a fashion that pores exist and that a substantial fraction of these pores are of dimensions of between 1 and 1000 nm. By selecting certain polymerization conditions, the hydrogel may be obtained in the form of an amorphous gel or as beaded resin. Such soft beads may have a diameter of between 1 and 1000 micrometer.

Hydrogels may be synthesized from the polymers and copolymers listed above and physically cross-linked or chemically cross-linked by radical, anionic or cationic polymerization, by chemical reactions like condensation or addition reactions as described in W.E. Hennink and C.F. van Nostrum, Adv. Drug Del. Rev. 2002, 54, 13-36.

Further examples include branched and hyperbranched polymers. Examples for such polymers include dendrimers and other dense star polymers. (R. Esfand, D.A. Tomalia, Drug Discov Today, 2001, 6(8), 427-436; P.M. Heegaard, U. Boas, Chem. Soc. Rev. 2004 (33(1), 43-63; S.M. Grayson, J.M. Frechet, Chem. Rev. 2001, 101 (12), 3819-3868).

R1 can also be a biopolymer like a protein. Non-limiting examples of such polymers include albumin, antibodies, fibrin, casein, and other plasma proteins.

Each R1 polymer can carry one or more biologically active substances linked to the polymer by conjugation with a second prodrug linker as described herein or any other linker known to the person skilled in the art. The polymers may have further substituents and may be functionalized for attachment to the spacer moiety X. Non-limiting examples of such functional groups comprise carboxylic acid and activated derivatives, amino, maleimide, thiol, sulfonic acid and derivatives, carbonate and derivatives, carbamate and derivatives, hydroxyl, aldehyde, ketone, hydrazine, isocyanate, isothiocyanate, phosphoric acid and derivatives, phosphonic acid and derivatives, haloacetyl, alkyl halides, acryloyl, arylating agents like aryl fluorides, hydroxylamine, disulfides like pyridyl disulfide, vinyl sulfone, vinyl ketone, diazoalkanes, diazoacetyl compounds, epoxide, oxirane, and aziridine.

Preferred functional groups for the R1 polymer include but are not limited to thiol, maleimide, amino, carboxylic acid and derivatives, carbonate and derivatives, carbamate and derivatives, aldehyde, and haloacetyl.

Especially preferred functional groups include thiol, maleimide, amino, carboxylic acid and derivatives, carbamate and derivatives, and carbonate and derivatives thereof.

Non-limiting examples for suitable bonds or groups formed between X and R1 include disulfide, S-succinimido, amide, amino, carboxylic ester, sulfonamide, carbamate, carbonate, ether, oxime, hydrazone, urea, thiourea, phosphate, phosphonate, etc.

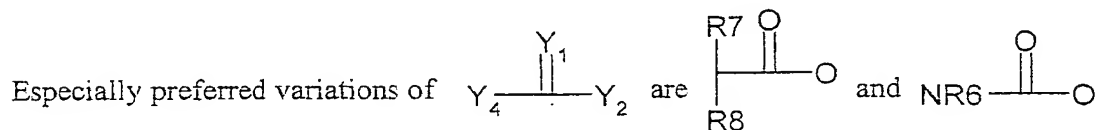
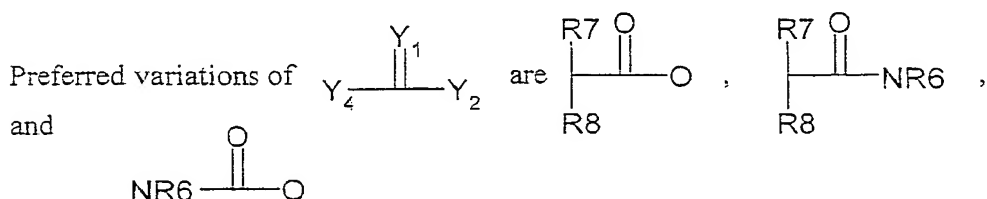
Preferred bonds or groups formed between X and R1 comprise S-succinimido, amide, carbamate, and urea.

Preferably, the R1 polymers are well hydrated, degradable or excretable, nontoxic and non-immunogenic in mammals. Preferred R1 polymers include polyalkoxy-based polymers like polyethylene glycol and polyethylene glycol reagents as those

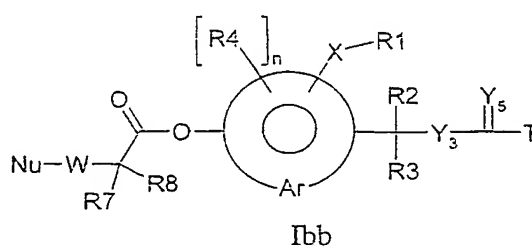
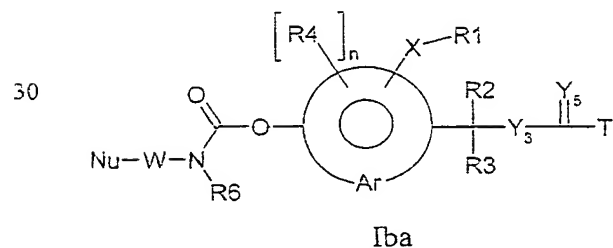
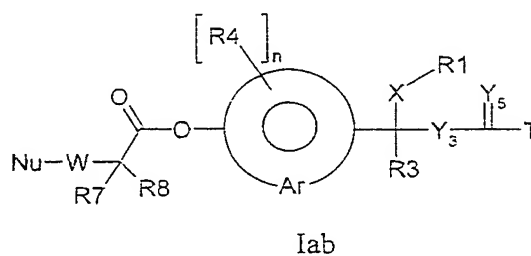
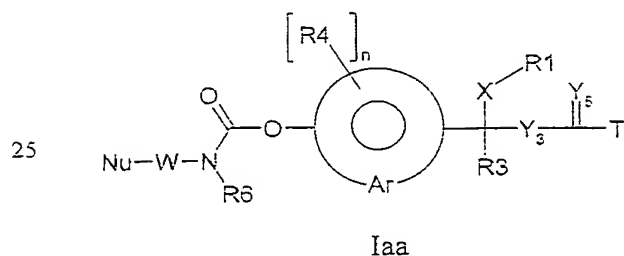
described in Nektar Inc. 2003 catalog "Nektar Molecule Engineering – Polyethylene Glycol and Derivatives for Advanced PEGylation" and branched, hyperbranched, cross-linked polymers and hydrogels, and proteins like albumin.

- 5 W is selected from substituted or non-substituted linear, branched or cyclical alkyl, aryls, substituted aryls, substituted or non-substituted linear, branched or cyclical heteroalkyl, substituted or nonsubstituted heteroaryls, etc.

W is selected preferably from non-toxic substituted or non-substituted linear,
10 branched or cyclical alkyls or heteroalkyls.



- 20 forming polymeric prodrugs of the following formulas:



In formula Ia and Ib R6 may also be Nu-W.

At least one Nu is present in Nu-W.

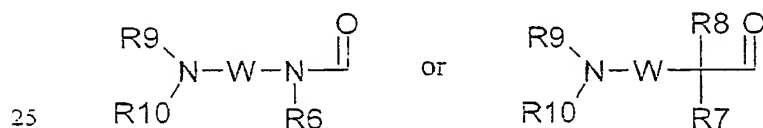
- 5 Nu is a nucleophile that can perform a nucleophilic attack at the carbonyl carbon of $\text{Y}_4-\overset{\text{Y}_1}{\underset{\text{Y}_2}{\text{C}}}=\text{O}$ and thus catalyse the cleavage of the masking group by intramolecular catalysis or cyclization (figure 8). Figure 8 shows an example according to formula Ia or Ib wherein the cleavage of the masking group is by intramolecular cyclization. In cases where Nu only catalyses the cleavage of the masking group by intramolecular
- 10 catalysis, no cyclical product of the masking group is formed.

Preferred nucleophiles include primary, secondary and tertiary amino groups, thiol, carboxylic acid, hydroxylamine, hydrazine, and nitrogen containing heteroaryl. Especially preferred nucleophiles include primary, secondary and tertiary amino groups. In order to effectively catalyse the cleavage of the masking group, the spacing

15 between the nucleophile Nu and Y₂ is preferably between three and fifteen atoms. More preferably, the spacing between Nu and Y₂ is between four and ten atoms. The at least one nucleophile Nu may be attached anywhere to W (e.g. at the terminus or in the middle of W) or may be part of W.

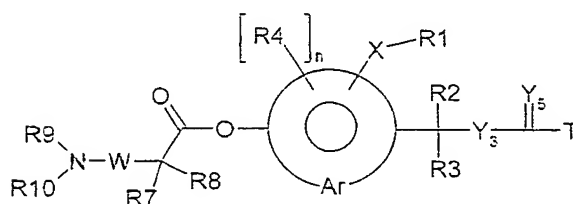
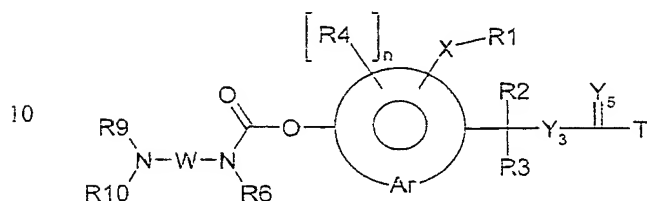
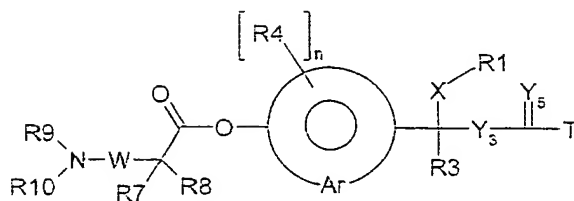
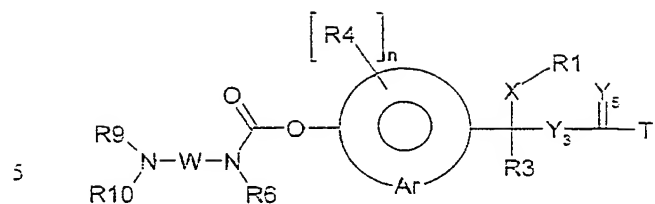
- 20 Preferred variations for the masking group $\text{Nu}-\text{W}-\text{Y}_4-\overset{\text{Y}_1}{\underset{\text{Y}_2}{\text{C}}}=\text{O}$

are selected independently from



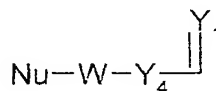
where $\begin{array}{c} \text{R}_9 \\ \diagdown \\ \text{N}-\text{W} \\ \diagup \\ \text{R}_{10} \end{array}$ forms a primary, secondary or tertiary amine nucleophile Nu.

- These preferred variations result in polymeric prodrugs according to the following
- 30 formulas:



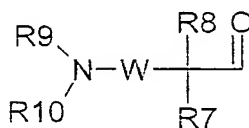
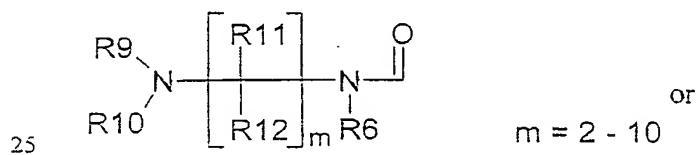
- 15 R9, R10 are selected independently from hydrogen, substituted or non-substituted alkyl or heteroalkyl, substituted or non-substituted aryl or heteroaryl.

Especially preferred variations for the masking group

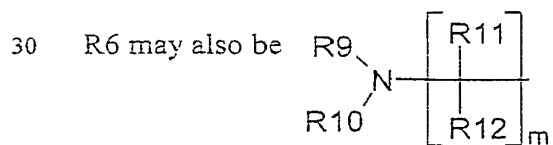


20

are selected from



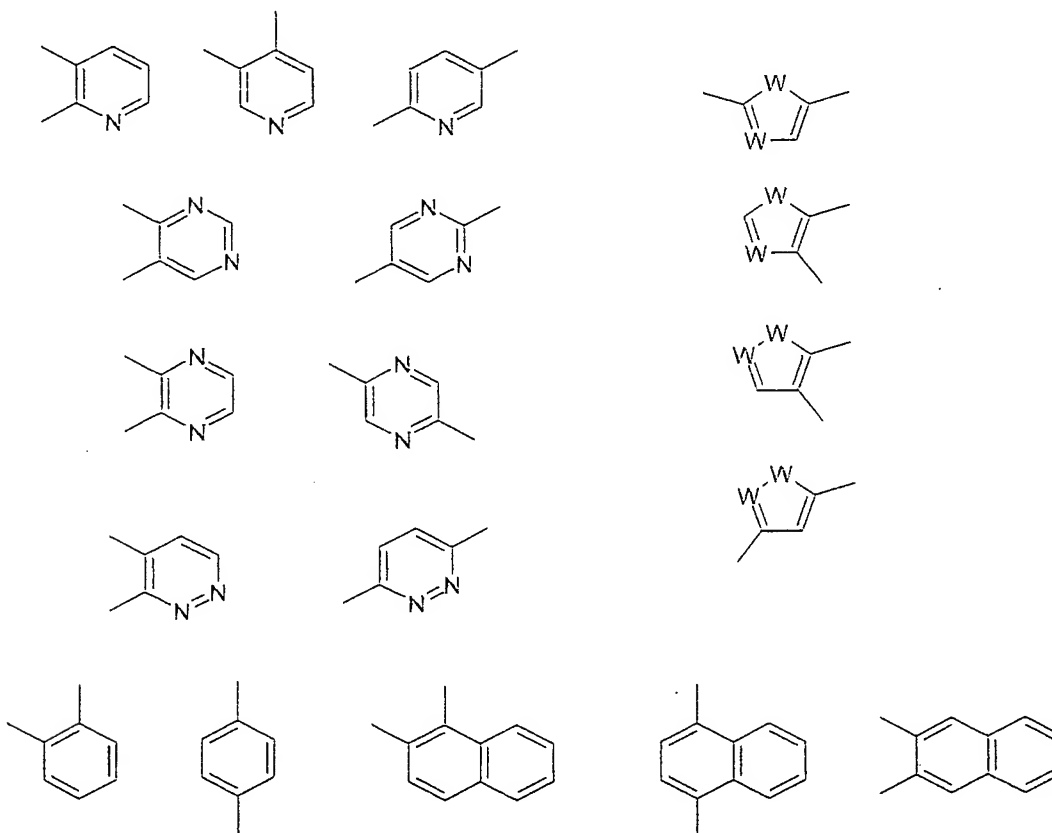
Preferably, R9, R10, R11 and R12 are selected independently from hydrogen, substituted or non-substituted alkyl and R7 and/or R8 are not hydrogen.



and is preferably not hydrogen.

Surprisingly it was found, that the masking group can modify irreversibly the amine containing biologically active moiety when the nucleophile Nu is absent in the masking group. As shown in the example section, during release of the bioactive moiety insulin from a polymeric prodrug with a pentanoyl masking group which is not part of the present invention (as it does not contain a nucleophile), approximately 30% of the insulin molecule was modified with the masking group by acyl transfer. The mechanism of an example of this modification where D contains an additional free amino group that serve as nucleophile for acyl transfer from the masking group is shown in figure 9.

Ar of formula Ia or Ib is a multi-substituted aromatic hydrocarbon or a multi-substituted aromatic heterocycle. To be aromatic, the number of pi electrons must satisfy the Hückel rule ($4n+2$) and the cycle has to be planar. A huge variety of compounds satisfy these criteria and thus are suitable as Ar in formula Ia or Ib. Non-limiting preferred aromatic moieties include:



wherein W is O, N, or S, independent from each other.

5 Y_2 and $\begin{array}{c} R1 \\ | \\ X \\ | \\ \text{---} Y_3 \text{---} \text{---} Y_5 \text{---} T \\ | \\ R3 \end{array}$ in formula Ia or Y_2 and $\begin{array}{c} R2 \\ | \\ \text{---} Y_3 \text{---} \text{---} Y_5 \text{---} T \\ | \\ R3 \end{array}$ in formula Ib have to be arranged on the aromatic ring in such a fashion that a 1,4- or 1,6- or 1,(4+2p), with p= 2, 3, 4 and higher, elimination can take place (see above). For example, in the case of a 6-membered ring, the substituents have to be arranged *ortho* or *para*.

10

Preferred moieties for Ar are mono- and dicyclic aromatic hydrocarbons or aromatic heterocycles.

Especially preferred moieties are monocyclic five- or six-membered aromatic
15 hydrocarbons or aromatic heterocycles.

General synthesis procedures of the polymeric prodrugs

Synthesis of representative examples of polymeric prodrugs according to the present
20 invention is described in the Examples section.

Prodrugs of the present invention can be prepared in various different fashions. Figure 10 shows general routes for the synthesis of the polymeric prodrugs of the present invention according to formula Ia.

25 In a first method, intermediate (III) is provided by acylating Y_2 of starting material (II) with the masking group $\begin{array}{c} Y_1 \\ | \\ Nu-W-Y_2 \end{array}$.

For this, X or Nu may have to be protected with a reversible protecting group PG₁. Suitable protecting groups are described in TW Greene, P.G.M. Wuts, Protective
30 groups in organic synthesis, 1999, John Wiley & Sons, 3rd ed.. From intermediate (III) two alternative routes can be used to yield (Iaa). In a first route intermediate (III) is activated by an activating agent such as 4-nitrophenyl chloroformate or disuccinyl carbonate to yield (IV). The amine containing drug molecule is attached to (IV) to yield (V) by displacing the leaving group of the activated intermediate (IV). After

deprotection of X such as by treating intermediate (V) with reagents like trifluoroacetic acid or DTT (where applicable) deprotected intermediate (V) is then reacted with polymer R1 to yield the polymeric prodrug (Iaa).

- 5 In a second route the polymer R1 is first attached to the intermediate (III) after deprotection of X (where applicable) to form intermediate (VI). After an activation step intermediate (VII) is formed. (VII) is reacted with the amine containing drug molecule to form the polymeric prodrug (Iac).
- 10 In a second method, intermediate (VIII) is provided by activating starting material (II) by an activating agent such as 4-nitrophenyl chlorofomate. For this, Y₂ and/or X may have to be protected with a protecting group PG₂ and/or PG₁. Amine containing drug is reacted with intermediate (VIII) to form (IX). In a first route, Y₂ of (IX) is selectively deprotected and acylated to form intermediate (V) which is further
- 15 processed to (Iaa) as described above. In a second route X is selectively deprotected and reacted with polymer R1 to form intermediate (X). Y₂ of (X) is then deprotected and acylated to form the polymeric prodrug (Iac).

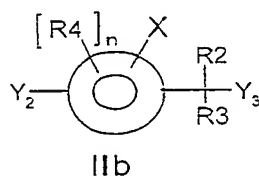
In a third method starting material (II) is reacted with polymer R1 to form

20 intermediate (XI). In one route, intermediate (XI) can be acylated to form intermediate (VI) which processed as described above to form polymeric prodrug (Iaa). In a second route, Y₂ is protected by the protecting group PG₂, activated and reacted with the amine containing drug molecule to form (X). Intermediate (X) is then processed as described above to form the polymeric prodrug (Iac).

25

For all methods described, further functional groups such as Y₃ or nucleophiles present in Nu-W may have to be protected with suitable protecting groups.

30 Polymeric prodrugs according to formula Ib can be prepared by methods described above for prodrugs according to formula Ia using starting material IIb instead of II in figure 10.



It is understood, that linker structures according to the outlined invention and carrying protecting groups or leaving groups as described and used in the synthesis of corresponding polymeric prodrugs are considered within the range of the invention.

5

Application of the polymeric prodrugs in molecular therapy

For polymeric cascade prodrugs it is desirable for the cleavage kinetics of the first temporary linkage to proceed under conditions present in the blood circulation of the human body (pH 7.4, 37°C). Most importantly, cleavage of the first temporary linkage should be based on hydrolysis and exhibit none or only very limited dependence upon chemical or biochemical or physicochemical entities present in the human blood circulation such as enzymes, salts or binding proteins.

15 It was now surprisingly found that the rate of cleavage of the first temporary linkage connecting the masking group with the activating group as well as its dependence upon blood components can be controlled by neighbouring group effects mediated by nucleophilic functional groups (such as primary, secondary or tertiary amines) present in the masking group and positioned in a distance to the corresponding temporary
20 bond. If the masking group is structured in this fashion, an intramolecular reaction with contribution of the nucleophile governs the kinetics of the linkage (Fig. 5 and Fig. 8).

Key advantage of the polymeric prodrugs of the present invention is their
25 predominantly non-enzymatic cleavage: the half-life of the prodrug in suitably buffered human blood plasma of pH 7.4 (with aqueous buffer concentration < 50 %) is at least 50 % of the half-life of the prodrug in enzyme-free buffer pH 7.4.

This feature allows for better predictability and control of release rates after
30 administration to a living organism and reduces interpatient variability.

In contrast to the enzymatic dependency of masking group removal as described in abovementioned examples of Antczak et al., Shabat et al. and Lee et al., a higher

level of control over release rates can be achieved if the masking group has enzyme-independent, self-eliminating properties.

The masking groups according to the present invention contain at least one
5 nucleophile Nu. Structural features of this masking group such as nucleophilicity of the amine group and ring-forming capacity may be systematically optimized in order to precisely adjust the rate of prodrug cleavage. Such intramolecular reactions resulting in unmasking and subsequent rearrangement are highly independent from enzymes due to the fact that intramolecular reactions are generally preferred over
10 intermolecular reactions as shown diagrammatically in Fig. 8.

In another embodiment of the invention, independency of prodrug cleavage from enzyme levels is achieved by providing a prodrug containing a sterically demanding carrier group as is shown in Fig. 7.

15 Such encapsulation or sterical protection by the sterically demanding carrier group may be conferred by a branched, hyperbranched, crosslinked or self-assembled structure of the carrier polymer. Such polymers tend to form a densely packed molecular volume, as exemplified for instance in dendrimers, dense star polymers or
20 bead-shaped nano- and microparticles or amorphous gels. If the linkage of the polymer carrier to the drug is located in the interior of the polymer carrier, the linked drug will be efficiently encapsulated and protected from enzymatic attack. In this case, sterical hindrance by the polymer prevents enzymes from accessing and cleaving the temporary linkages.

25 In yet another embodiment, enzyme-independent prodrug cleavage is achieved by combining an intramolecular self-eliminating masking group with an encapsulating hyperbranched or crosslinked or self-assembled carrier.

30 A further advantage of the present invention is the release of an unmodified biologically active moiety. In cases where the biologically active moiety contains further reactive functional groups like amino groups of lysine residues in proteins, an unwanted side reaction between the masking group and the biologically active moiety can occur. The reactive functional groups of the biologically active moiety may react

with the masking group, forming a stable covalent bond and resulting in the release of a modified biologically active moiety. This potential side reaction is shown schematically in Figure 9. The occurrence of such side reactions is shown in the examples section using polymeric prodrugs which are not part of the present invention with simple masking groups like a pentanoyl residue without a nucleophile Nu present in the masking group and as described by Antczak et al. or Lee et al. The side reaction in this linker system is suppressed using polymeric prodrugs according to the present invention with intramolecularly activated masking groups that contain nucleophiles Nu (see example section).

Enzyme-independent release control enables depot formulations without the need for encapsulation. Until now, many biocompatible materials like hydrogels with large pore sizes could not be used for depot formulations due to their lack of encapsulation properties. From such well-hydrated and mechanically soft biocompatible materials, biologically active moiety would be released too fast for most therapeutic applications. In combination with the prodrug linkers described in this invention, the carrier material may be optimized for its biocompatibility properties as the release is solely governed by the linker cleavage kinetics and does not require chemical or enzymatic degradation of the polymer carrier itself.

Release rates are governed by a substantially non-enzymatic chemical reaction which is in turn dependent on the molecular structure of the linker. Systematic or random modifications of the chemical structure, for instance by changing substituents in one or more positions, for instance a masking group in a cascade prodrug, allows for the generation of prodrug linkers with differing release rates. It is therefore possible to create a variety of prodrug linkers and select those fast or slow cleaving prodrug linkers according to the demands posed by a given medicinal or therapeutic application.

Another advantageous feature which is part of this invention is the attachment of the polymer carrier through a stable covalent bond to an activating moiety involved in a double or cascade prodrug release mechanism. As part of this invention, the activating moiety remains attached to the polymer carrier after drug release and therefore cannot diffuse into the environment. Permanent linkage of the polymer carrier to the activating

group greatly reduces the any side-reactivity of the activating moiety and the probability of unwanted toxic effects. In other polymeric cascade prodrugs known in the art, the activating moiety is released in addition to the drug. Due to the molecular rearrangement mechanisms used in cascade prodrugs, the activating moiety is released
5 in a highly reactive form and may cause direct damage to surrounding biomolecules, or potentially toxic derivatives of the activating moiety may be formed in vivo.

Description of the Figures

- 10 Fig. 1 shows a carrier-linked prodrug.
Fig. 2 shows an enzyme-dependent carrier-linked prodrug.
Fig. 3 shows a cascade prodrug where the masking group is part of the carrier.
Fig. 4 shows an enzyme-dependent cascade prodrug where the masking group is part
of the carrier.
15 Fig. 5 shows a self-cleaving cascade prodrug where the masking group is separate
from the carrier.
Fig. 6 shows an enzyme-dependent cascade prodrug where the masking group is
separate from the carrier.
Fig. 7 shows a cascade prodrug where the carrier is sterically protecting the masking
20 group.
Fig. 8 shows cleavage of the masking group by intramolecular cyclisation.
Fig. 9 shows a possible side reaction of polymeric prodrug activation.
Fig.10 shows general synthesis methods.
Fig.11 shows mass spectra of prodrug released insulin molecules.

Examples

Materials

Fmoc-amino acids, resins and PyBOP were purchased from Novabiochem and are
5 named according to the catalogue. Fmoc-Ado-OH was obtained from Neosystem. All
additional chemicals were purchased from Sigma Aldrich. Recombinant human
insulin was from ICN Biomedicals (USA). Maleimide-PEG5k was obtained from
Nektar (USA). 5-(and-6)-carboxyfluorescein succinimidyl ester (mixed isomers) was
obtained from Molecular Probes.

10

Solid phase synthesis reaction medium

Solid phase synthesis was performed on NovaSyn TG Sieber amide resin with a
loading of 0.17 mmol/g or 2-chlorotrityl chloride resin with a loading of 1.4 mmol/g.
Syringes equipped with polypropylene frits were used as reaction vessels.

15

Standard coupling cycle for fmoc-protected amino acids

For fmoc protecting-group removal, the resin was repeatedly (three times, 4 min each)
agitated with 2/2/96 (v/v/v) piperidine/DBU/DMF and repeatedly (six times) washed
with DMF.

20

Coupling of fmoc-protected amino acids to free amino groups on resin was achieved
by agitating the resin with 3 equivalents (eq) of fmoc-amino acid, 3 eq PyBOP and 6
eq DIEA in relation to free amino groups in DMF for 60 min.

Finally, the resin was repeatedly (five times) washed with DMF.

25

Standard cleavage protocol for TentaGel Sieber amide resin

Upon completed synthesis, the resin was washed with DCM, dried in vacuo and
treated repeatedly (five times) with 97/2/1 (v/v) DCM/TES/TFA. After evaporation,
compounds were purified by preparative RP-HPLC (Waters 600).

30

Standard cleavage protocol for 2-chlorotrityl chloride resin

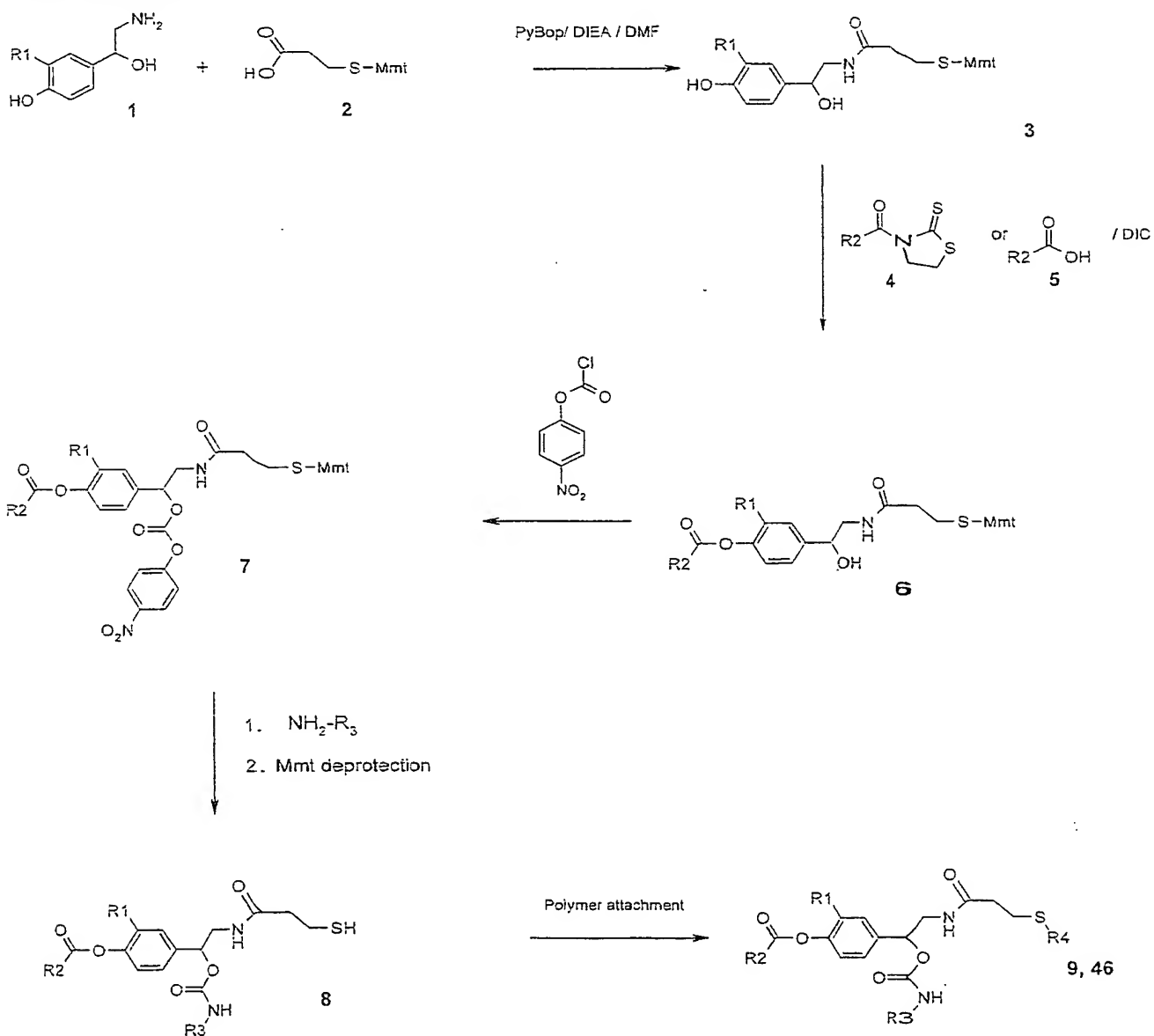
Upon completed synthesis, the resin was washed with DCM, dried in vacuo and
treated two times for 30 minutes with 65/35 (v/v) HFIP/DCM. After combining the
eluates the volatile components were evaporated.

Analysis

Mass spectrometry (MS) was performed on a Waters ZQ 4000 ESI instrument and spectra were, if necessary, interpreted by Waters software MaxEnt.

- 5 Size exclusion chromatography was performed using an Amersham Bioscience AEKTA basic system equipped with a Superdex 200 column (Amersham Bioscience). NMR spectra were recorded on a Bruker AC300.

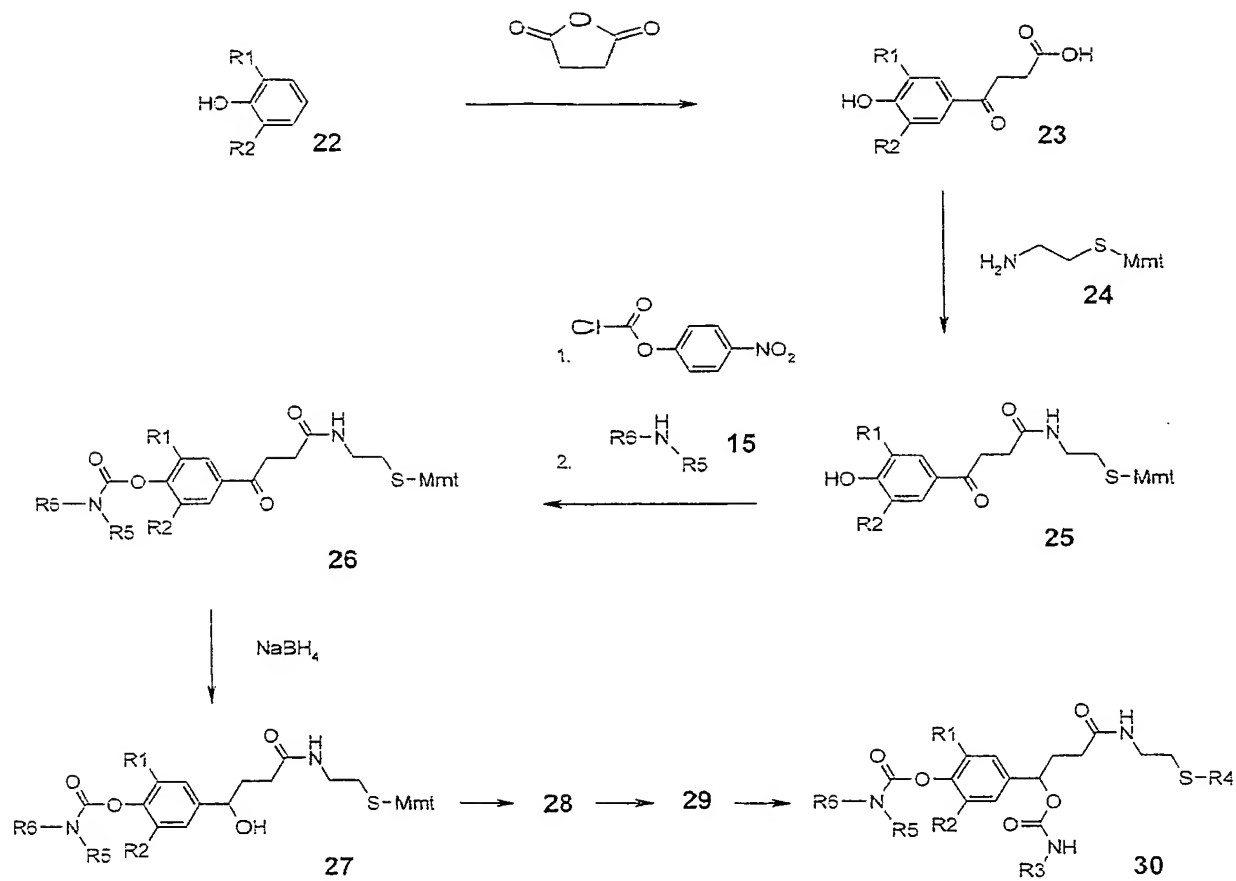
10 Overview - synthesis of polymeric prodrugs according to formula Ia with ester-linked masking groups



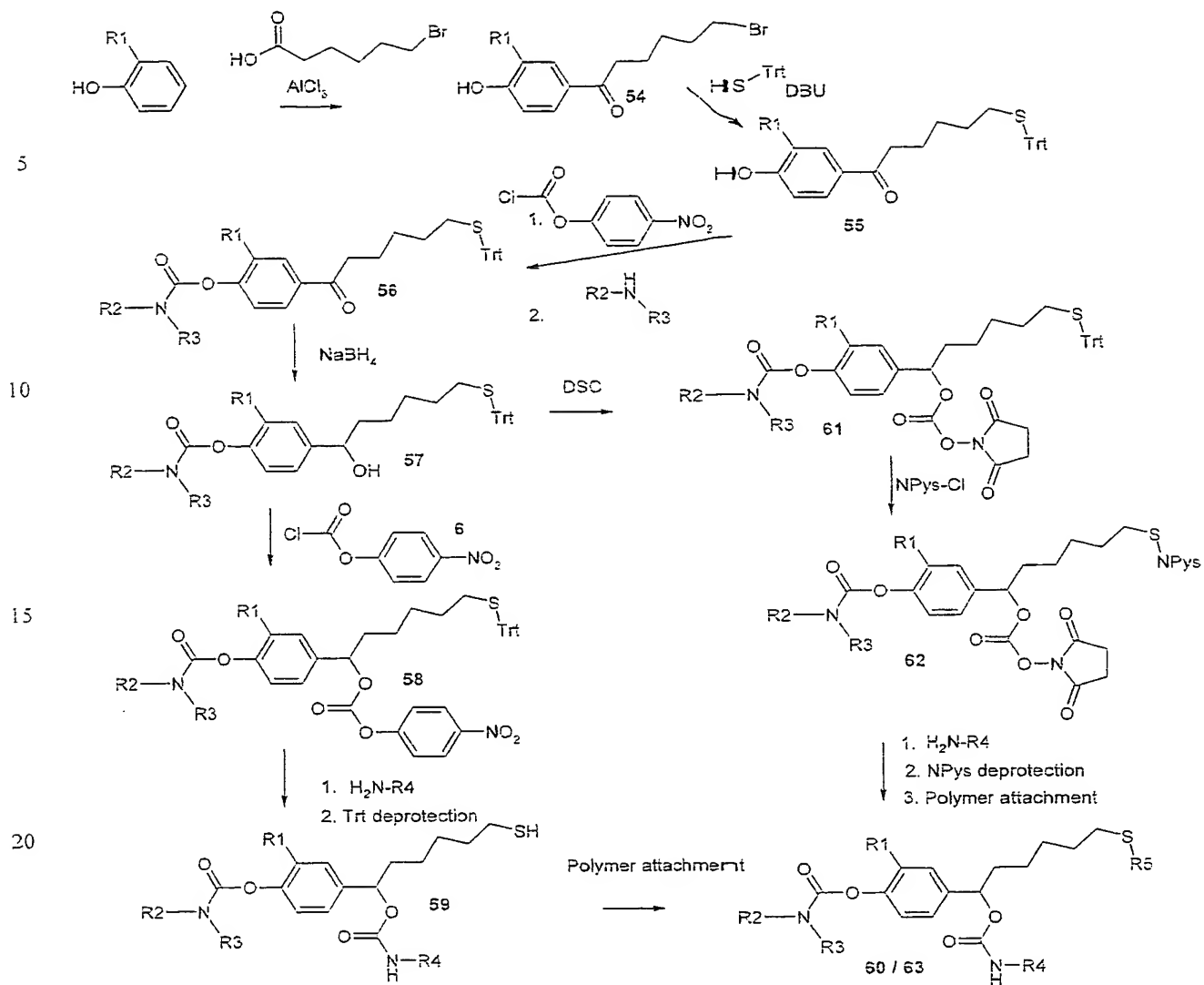
2)



b)

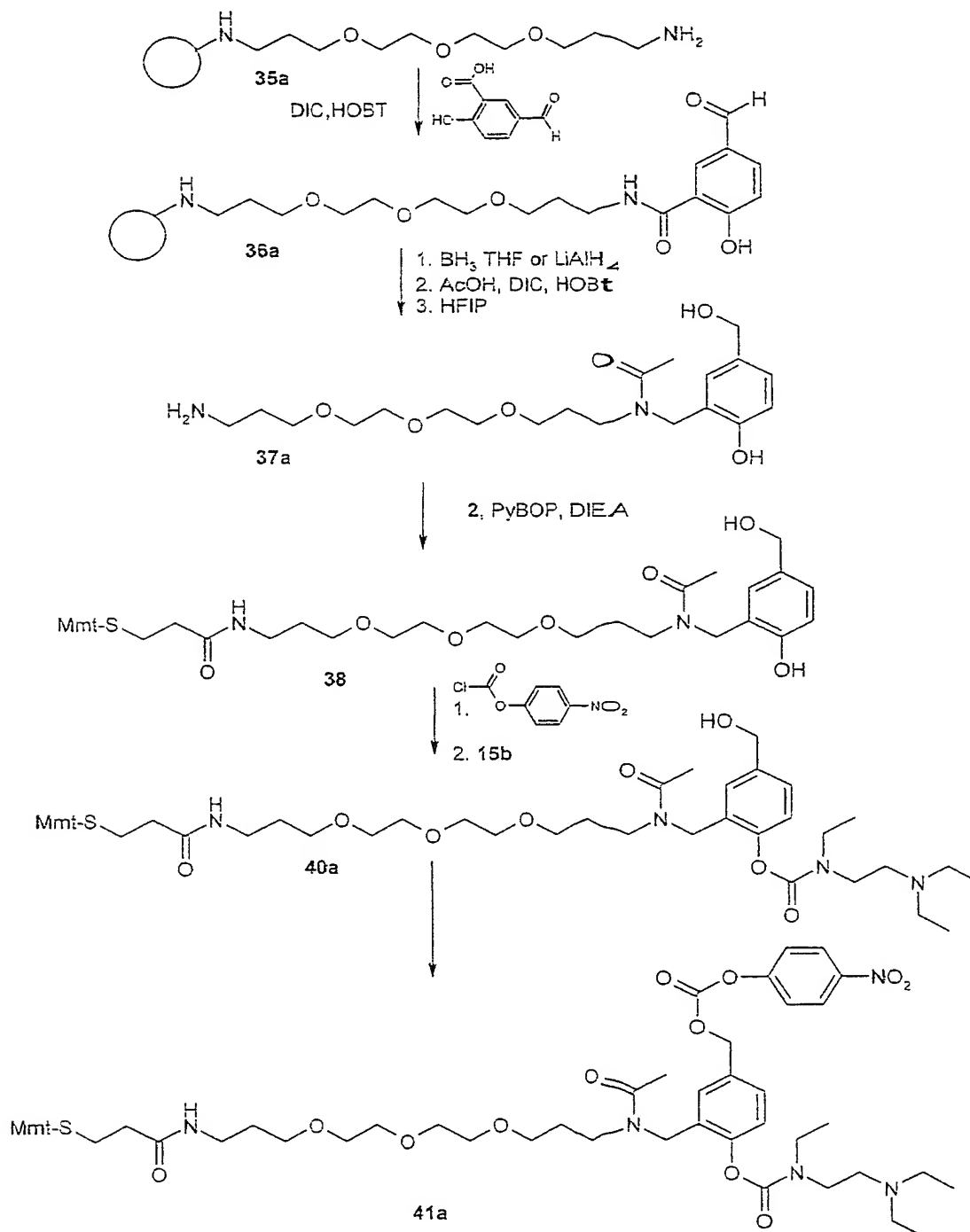


c)

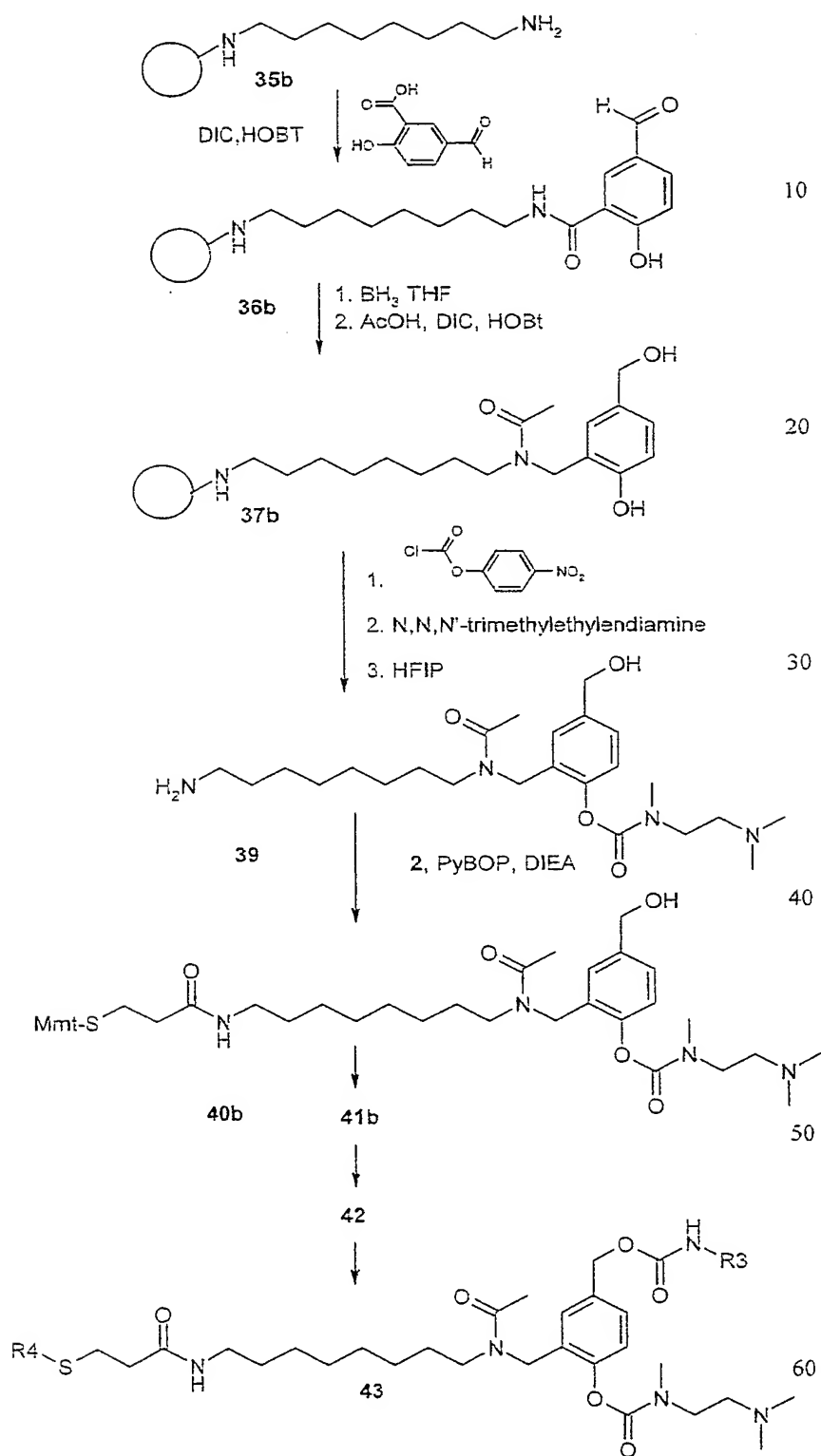


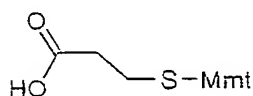
Synthesis of polymeric prodrugs according to formula Ib with a carbamate-linked masking group

a)

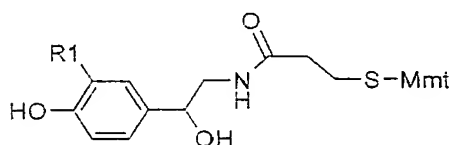


50 b)



Synthesis of compound 2**2**

Mmt-chloride (1 eq) and mercaptopropionic acid (1.1 eq) were dissolved in TFA and incubated for 30 min. The solvent was removed under reduced pressure. The product was dissolved in pyridine, diluted in water, acidified by acetic acid and extracted with ether. The ether phase was separated and dried over Na₂SO₄. Solvent was removed under reduced pressure and product **2** was purified by RP-HPLC.

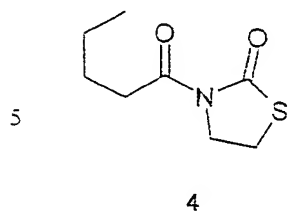
Synthesis of compounds 3a and 3b**3a** R1 = H**3b** R1 = OMe

Octopamine hydrochloride (**1a**) (2 eq), DIEA (4 eq), and PyBOP (1 eq) were dissolved in DMF, **2** (1 eq) was added and the mixture was reacted for 50 min at room temperature. After addition of acetic acid (7 eq) product **3a** was purified by RP-HPLC.

3b was synthesized from normetanephrine hydrochloride (**1b**) as described above.

3a: MS [M+Na]⁺ = 536 (MW+Na calculated = 536.2 g/mol)

3b: MS [M+Na]⁺ = 566 (MW+Na calculated = 566.2 g/mol)

Synthesis of mercaptothiazolide 4

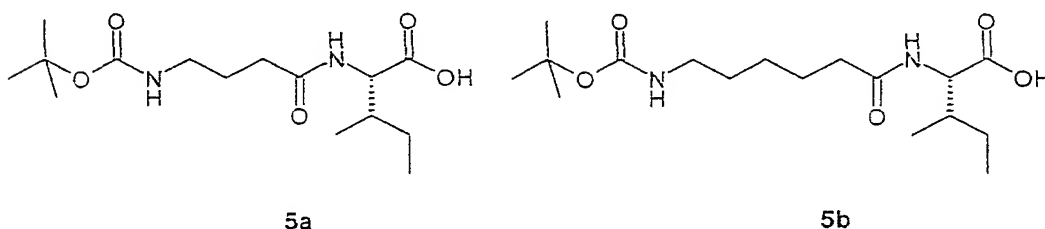
2-Mercaptothiazoline and triethylamine (1.5 eq) were dissolved in dry THF and pentanoyl chloride (1 eq) was added. The mixture was stirred for 1 h at 50 °C under
 10 inert gas atmosphere and was allowed to cool to room temperature. 0.5 N aqueous HCl was added and the separated organic phases were dried over Na₂SO₄. After concentration *in vacuo* the residue was purified by silica gel column chromatography using heptane/ethyl acetate (1/1) as mobile phase. Mercaptothiazolide 4 was collected as a viscous yellow oil.

15

4 R_f (heptane/ethyl acetate 1:1) = 0.7

Synthesis of compounds 5a and 5b

20



25

General synthesis protocol:

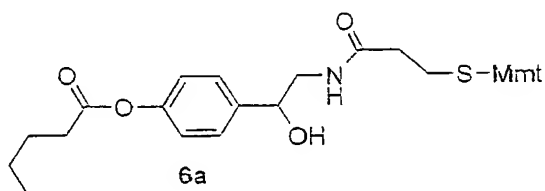
1 g 2-chlorotrityl chloride resin (loading 1.6 mmol/g) was incubated for 1 h with 850 mg (2.4 mmol) Fmoc-Ile-OH and 840 µl (4.8 mmol) DIEA in 5 ml 1/1 DCM/DMF. After fmoc removal and washing of the resin with DMF, boc-aminobutyric acid was
 30 coupled to 0.5 g resin according to the standard coupling method. Compound 5a was cleaved from the resin with 97/1/2 (v/v) DCM/TFA/TES for 45 min. After neutralisation with pyridine, solvents were removed under reduced pressure and 5a was purified by RP-HPLC.

5b was synthesized from boc-aminohexanoic acid as described above.

5a MS $[M+Na]^+ = 339.2$ (MW+Na calculated = 339.4 g/mol)

5b MS $[M+Na]^+ = 367.4$ (MW+Na calculated = 367.5 g/mol)

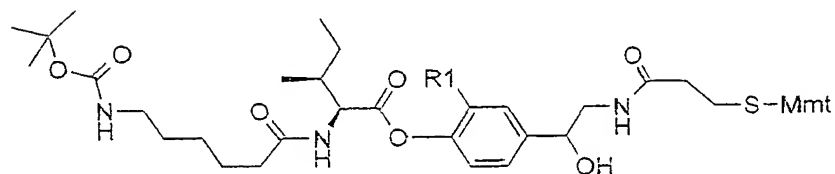
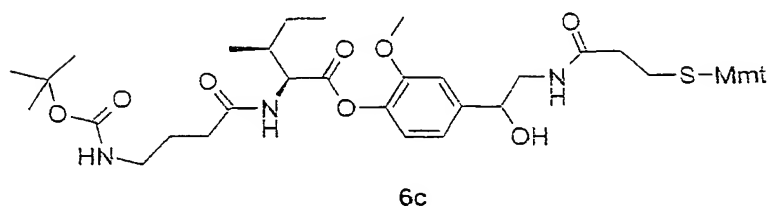
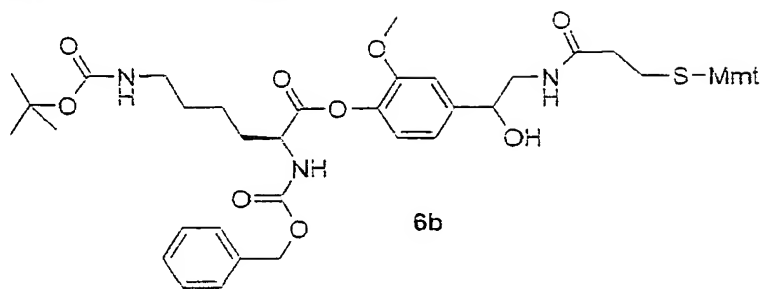
5 Synthesis of compound 6a



Mercaptothiazolide 4 (1 eq), phenol 3a (4 eq) and DMAP (4 eq) were refluxed in DCM for 2 h under nitrogen atmosphere. After neutralization with acetic acid, the solvent was removed *in vacuo* and product 6a was purified by RP-HPLC.

6a MS $[M+Na]^+ = 620$ (MW+Na calculated = 620.3 g/mol)

Synthesis of compounds **6b** to **6e**



General synthesis protocol:

Carboxylic acid **5a** (1 eq), phenol **3b** (1 eq), DIC (1 eq), and DMAP (2 eq) in DMF were reacted for 1h at room temperature. After addition of acetic acid (4 eq) the resulting carboxylic ester **6c** was purified by RP-HPLC.

6d was synthesized as described above using **5b** and **3b** as starting materials.

6b was synthesized as described above using Z-Lys(Boc)-OH and **3b**.

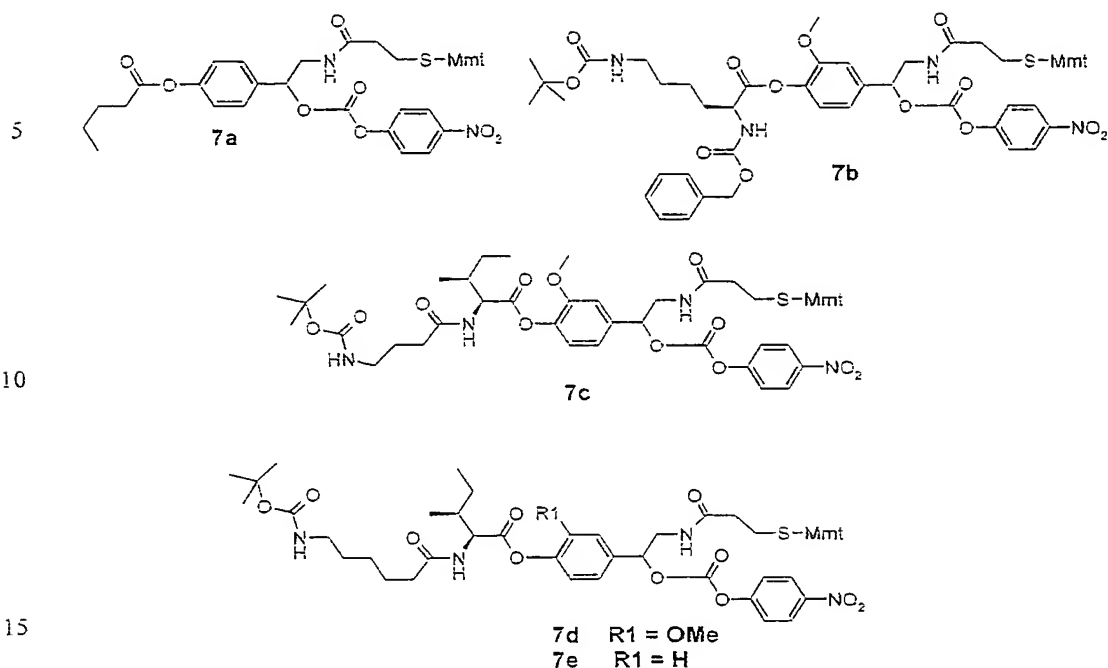
6e was synthesized as described above using **5b** and **3a**.

6b MS $[M+Na]^+ = 928$ (MW+Na calculated = 928.6 g/mol)

6c MS $[M+Na]^+ = 864$ (MW+Na calculated = 864.5 g/mol)

6d MS $[M+Na]^+ = 892$ (MW+Na calculated = 892.6 g/mol)

6e MS $[M+Na]^+ = 862$ (MW+Na calculated = 862.6 g/mol)

Synthesis of compounds 7a to 7e

General synthesis protocol:

Alcohol **6a** (1 eq), 4-nitrophenyl chloroformate (10 eq), and DIEA (10 eq) were stirred in dry dioxane for 3 h at room temperature under nitrogen atmosphere. After addition of acetic acid (25 eq) the mixtures were diluted with 7/3 (v/v) acetonitrile/H₂O and the carbonate **7a** was purified by RP-HPLC.

7b, **7c**, **7d**, or **7e** were synthesized from **6b**, **6c**, **6d**, or **6e**, respectively, as described above.

7a MS [M+Na]⁺ = 785 (MW+Na calculated = 785.5 g/mol)

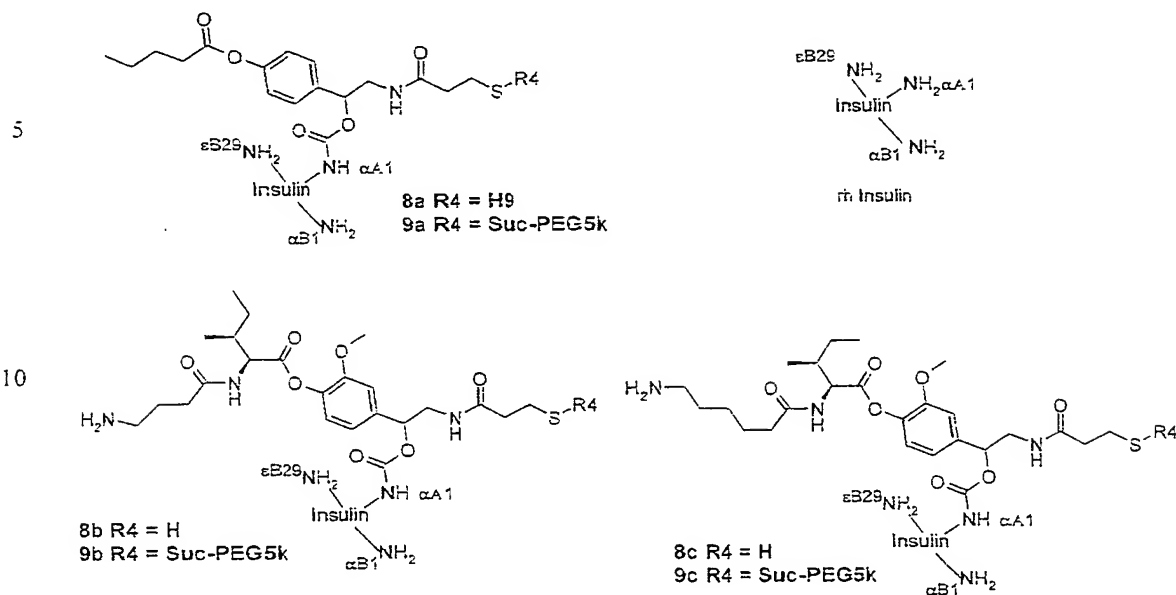
7b MS [M+Na]⁺ = 1093 (MW+Na calculated = 1093.7 g/mol)

7c MS [M+Na]⁺ = 1029 (MW+Na calculated = 1029.6 g/mol)

7d MS [M+Na]⁺ = 1057 (MW+Na calculated = 1057.6 g/mol)

7e MS [M+Na]⁺ = 1027 (MW+Na calculated = 1027.6 g/mol)

Synthesis of compounds 8a to 8c (N^{αA1}-linker-insulin)



General synthesis protocol:

Rh-Insulin in 1/1 (v/v) DMSO/DMF was mixed with a solution of 0.9 eq carbonate 7a in DMSO. The resulting solution was adjusted to basic pH with DIEA and stirred for 1.5 h at RT. RP-HPLC purification gave Mmt-protected intermediate.

After lyophilization, the Mmt-protected intermediate was mixed with 95:5 (v/v) TFA/triethylsilane and stirred for 5 min. Volatiles were removed under nitrogen flow and 8a was purified by RP-HPLC and lyophilized. Regioselectivity of insulin modification was verified by DTT reduction and MS analysis.

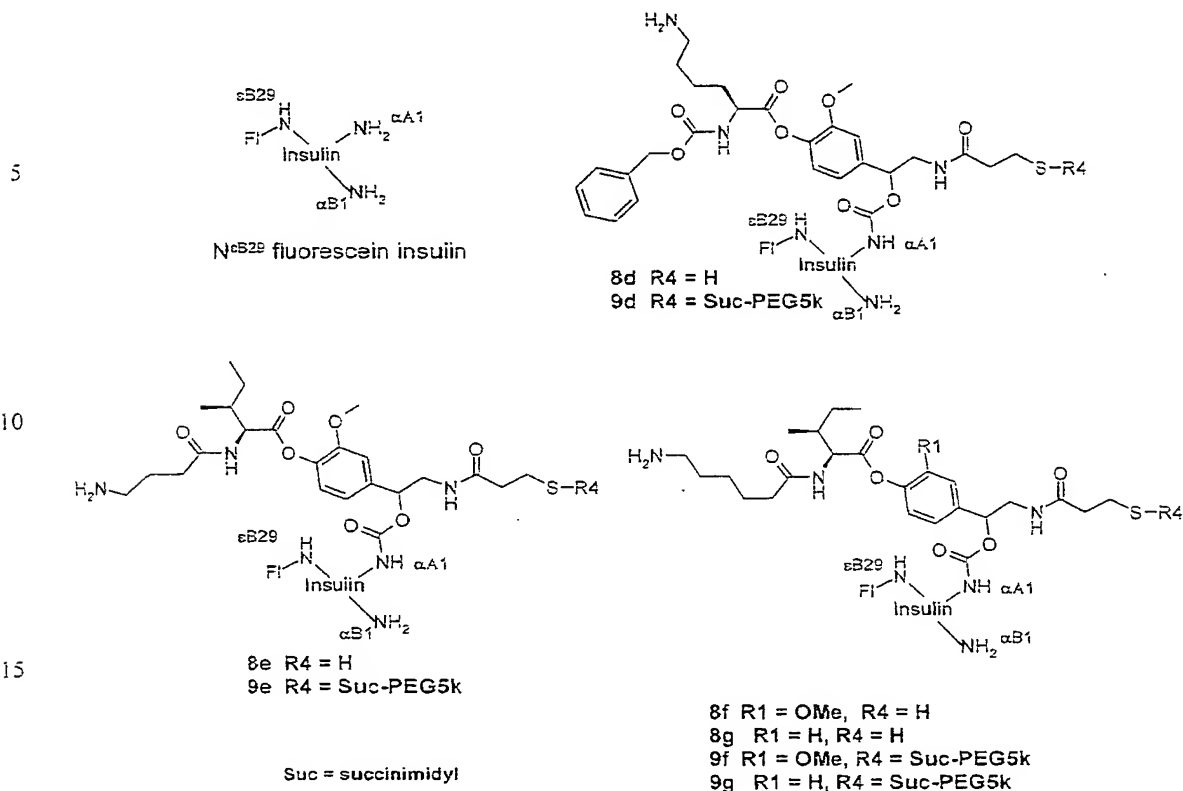
8b or 8c were synthesized from 7c, or 7d, respectively, as described above.

8a MS $[M+2H]^{2+} = 3078.9$; $[M+3H]^{3+} = 2053.2$ $[M+4H]^{4+} = 1540.6$ (MW calculated = 6158 g/mol)

8b MS $[M+2H]^{2+} = 3152.9$; $[M+3H]^{3+} = 2100.6$ $[M+4H]^{4+} = 1575.8$ (MW calculated = 6302 g/mol)

8c MS: $[M+3H]^{3+} = 2110.7$; $[M+4H]^{4+} = 1583.7$; $[M+5H]^{5+} = 1266.6$ (MW calculated = 6330 g/mol)

Synthesis of compounds 8d to 8g (N^{εB29}-Fluorescein-N^{αA1}-linker-insulin)



Synthesis of N^{εB29}-fluorescein insulin:

80 mg (13.8 μmol) rh-insulin were dissolved in 4 ml 1/1 (v/v) DMF/DMSO and 40 μl DIEA were added. 8 mg (17 μmol) 5-(and-6)-carboxyfluorescein succinimidyl ester were added and the solution was stirred for 30 min at room temperature. 4 ml 5/5/1 (v/v/v) acetonitrile/water/acetic acid were added, product N^{εB29}-fluorescein insulin was purified by RP-HPLC and lyophilized. The conjugation site was verified by reduction of N^{εB29}-fluorescein insulin with 1,4-dithiothreitol, protease digestion and MS analysis.

MS: [M+2H]²⁺ = 3084.0; [M+3H]³⁺ = 2054.6 (MW calculated = 6166 g/mol)

N^εB²⁹-fluorescein insulin in 1/1 (v/v) DMF/DMSO was mixed with a solution of 0.9 eq carbonate **7b** in DMSO. The resulting solution was adjusted to basic pH with DIEA and stirred for 3 h at RT. RP-HPLC purification gave Mmt-protected intermediate.

- 5 After lyophilization, the intermediate was dissolved in 95/5 (v/v) TFA/triethylsilane and stirred for 5 min. Volatiles were removed under nitrogen flow and **8d** was purified by RP-HPLC and lyophilized.

8e, **8f**, or **8g** were synthesized as described above using **7c**, **7d**, or **7e**, respectively.

- 10 **8d** MS: $[M+2H]^{2+} = 3364.1$; $[M+3H]^{3+} = 2242.7$; $[M+4H]^{4+} = 1681.5$ (MW calculated = 6724 g/mol)

8e MS: $[M+3H]^{3+} = 2219.2$ $[M+4H]^{4+} = 1665.9$; $[M+5H]^{5+} = 1332.8$ (MW calculated = 6660 g/mol)

- 8f** MS: $[M+3H]^{3+} = 2229.7$ $[M+4H]^{4+} = 1673.3$; $[M+5H]^{5+} = 1337.7$ (MW calculated = 6689 g/mol)

15 **8g** MS: $[M+3H]^{3+} = 2218.7$ $[M+4H]^{4+} = 1664.9$ (MW calculated = 6659 g/mol)

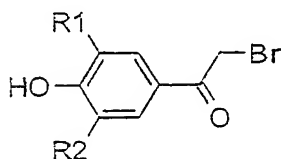
Synthesis of compounds **9a** to **9g** (mono-pegylated insulin compounds)

20

70 μ l 500 μ M **8a** in 1/4 (v/v) acetonitrile/water were mixed with 7 μ l 10 mM maleimide-PEG5k in 1/4 (v/v) acetonitrile/water and 10 μ l 0.5 M sodium phosphate buffer pH 7.0 and incubated for 15 min. Compound **9a** was purified by SEC (column: Superdex 200, flow rate: 0.75 ml/min) using 10 mM HEPES buffer (pH 7.4), 150 mM NaCl, 3 mM EDTA, and 0.005% Tween as mobile phase. The collected eluate (approximately 1.5 ml) was directly used as such for release rate determination.

25 **9b**, **9c**, **9d**, **9e**, **9f**, or **9g** were synthesized as described above from **8b**, **8c**, **8d**, **8e**, **8f**, or **8g**, respectively.

- 30 **9a** through **9g**: SEC retention time: 19.5 min

Synthesis of compounds 11a and 11b

11a R1 = R2 = Me

11b R1 = Me, R2 = H

10 3,5-Dimethyl-4-hydroxy-acetophenone (5.0 mmol) (**10a**) and CuBr₂ (1.7 g, 7.5 mmol) were dissolved in 10 ml ethyl acetate and refluxed for 2 h. Solid byproducts were removed by filtration. The filtrate was evaporated and crude product **11a** was purified by RP-HPLC.

11b was synthesized from 4-hydroxy-3-methyl-acetophenone (**10b**) (0.75 g, 5.0 mmol) as described above.

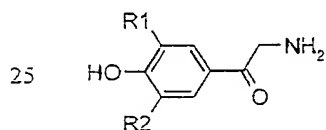
15

11a: Yield 754 mg (62%)

MS [M+H]⁺ = 243.1/245.1 (MW+H calculated = 244.1 g/mol)

11b: Yield 533 mg (47%)

20 MS [M+H]⁺ = 229.2/231.1 (MW+H calculated = 230.1 g/mol)

Synthesis of compounds 12a and 12b

12a R1 = R2 = Me

12b R1 = Me, R2 = H

30 500 mg **11a** (2.06 mmol) and 576 mg (4.11 mmol) hexamethylenetetramine were dissolved in 20 ml of trichloromethane and refluxed for 30 min. The solvent was removed in vacuo. 4 ml ethanol and 2 ml of concentrated HCl were added and the slurry was heated to 50 °C for 4 h. The mixture was concentrated in vacuo, diluted with acetonitrile/water and **12a** was purified by RP-HPLC.

12b was synthesized from 472 mg (2.06 mmol) 11b as described above.

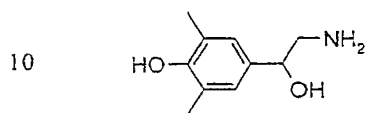
12a: Yield 547 mg (81%) as TFA-salt

MS $[M+Na]^+ = 202.2$ (MW+Na calculated = 202.2 g/mol)

5 12b: Yield 455 mg (70%) as TFA-salt

MS $[M+Na]^+ = 188.2$ (MW+Na calculated = 188.2 g/mol)

Synthesis of compound 13



13

500 mg (1.71 mmol) 12a (TFA salt) were dissolved in 10 ml 1/1 (v/v)

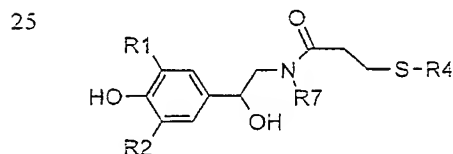
15 methanol/water, 129 mg (3.41 mmol) NaBH₄ were added and the mixture was stirred for 30 min at RT. 0.5 ml of acetic acid were added and 13 was purified by RP-HPLC.

13: Yield 313 mg (62%) as TFA-salt

MS $[M+Na]^+ = 204.2$ (MW+Na calculated = 204.2 g/mol)

20 NMR (300 MHz, DMSO-d₆) δ [ppm] = 8.25 (s, 1H, Phenol), 7.84 (bs, 3H, NH₃⁺), 6.89 (s, 2H, CH_{ar}), 5.85 (d, 1H, Hydroxyl, J=3.7 Hz), 4.62 (m, 1H, CH_{Benzyl}), 2.93 (m, 1H, CH₂), 2.80 (m, 1H, CH₂), 2.17 (s, 6H, CH₃).

Synthesis of compounds 14a to 14d



14a R1 = R2 = Me, R7 = H, R4 = Mmt

30 14b R1 = R2 = H, R7 = Me, R4 = Mmt

14c R1 = OMe, R2 = H, R7 = Me, R4 = Mmt

14d R1 = H, R2 = H, R7 = Me, R4 = Trt

13 (TFA salt, 159 mg, 0.541 mmol) was coupled to compound 2 as described for compound 3a to yield 14a.

14b, or 14c were synthesized as described above using synephrine (335 mg, 2.00 mmol) or metanephrine (HCl salt, 281 mg, 1.20 mmol), respectively.

5 Synephrine (335 mg, 2.3 mmol) was coupled to 3-tritylsulfanyl-propionic acid as described above to yield 14d.

14a: Yield 254 mg (87%)

MS $[M+Na]^+ = 564.7$ (MW+Na calculated = 564.3 g/mol)

10 14b: Yield 760 mg (72%)

MS $[M+Na]^+ = 550.2$ (MW+Na calculated = 550.3 g/mol)

14c: Yield 530 mg (80%)

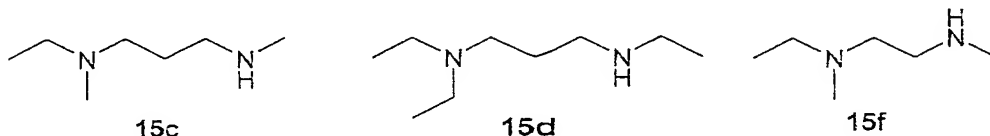
MS $[M+Na]^+ = 580.4$ (MW+Na calculated = 580.4 g/mol)

14d: Yield 567 mg (49%)

15 MS $[M+Na]^+ = 520.5$ (MW+Na calculated = 520.7 g/mol)

Synthesis of compounds 15c, 15d and 15f

20



General synthesis protocol:

25 1 g 2-chlorotriyl chloride resin (loading 1.4 mmol/g) was incubated for 1 h with N,N'-dimethylpropane-1,3-diamine (for synthesis of 15c), or N,N'-diethyl-propane-1,3-diamine (for synthesis of 15d) or N,N'-dimethyl-ethane-1,2-diamine (4 eq) (for synthesis of 15f) in DCM. After washing of the resin with DMF, amines were acetylated with 1/1/2 (v/v/v) acetic anhydride/pyridine/DMF for 14 h. The resin was washed with THF and dried. LiAlH₄ (1 M in THF, 4 eq) was added dropwise to the
30 suspended resin in THF. The resulting suspension was stirred for 3 h at 45°C under nitrogen atmosphere. After cooling, aqueous Rochelle's salt solution was added and the resin was separated and dried. Compounds were cleaved from resin with 2/1 (v/v)

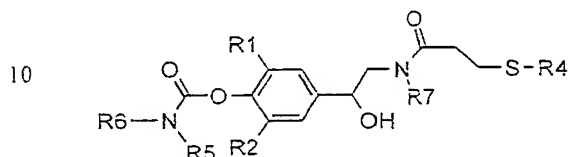
HFIP/DCM (2 x 30 min). The volatile components were evaporated and the products **15c**, **15d** or **15f** were used in the following steps without further purification.

15c MS. $[M+H]^+ = 131.2$ (MW = 130.1 g/mol)

5 **15d** MS $[M+H]^+ = 159.2$ (MW = 158.1 g/mol)

15f MS $[M+H]^+ = 117.1$ (MW = 116 g/mol)

Synthesis of compounds **16a** to **16f** and **16i**



16a R1 = R2 = R5 = Me, R6 = 2-(dimethylamino)ethyl, R7 = H, R4 = Mmt

16b R1 = OMe, R2 = H, R5 = Et, R6 = 2-(diethylamino)ethyl, R7 = H, R4 = Mmt

15 **16c** R1 = OMe, R2 = H, R5 = Me, R6 = 3-(N-ethyl-N-methylamino)propyl, R7 = Me, R4 = Mmt

16d R1 = R2 = H, R5 = Me, R6 = 3-(N-ethyl-N-methylamino)propyl, R7 = Me, R4 = Mmt

16e R1 = OMe, R2 = H, R5 = Et, R6 = 3-(diethylamino)propyl, R7 = Me, R4 = Mmt

20 **16f** R1 = R2 = H, R5 = Et, R6 = 3-(diethylamino)propyl, R7 = Me, R4 = Mmt

16i R1 = R2 = H, R5 = Et, R6 = 2-(diethylamino)ethyl, R7 = Me, R4 = Trt

25 **14a** (120 mg, 0.222 mmol) was dissolved in 1.5 ml of dry THF. p-Nitrophenyl-chloroformate (45 mg, 0.222 mmol) and DIEA (113 μ l, 0.665 mmol) were added and the mixture was stirred for 30 min at RT. **15a** (N,N,N'-trimethyl-ethylene-1,2-diamine) (72 μ l, 0.554 mmol) was added and stirring was continued for 30 min. The solvent was removed in vacuo, 100 μ l of AcOH were added and **16a** was purified by RP-HPLC.

30 **16b** was synthesized as described above from **3b** (80 mg, 0.15 mmol) and **15b** (N,N,N'-triethyl-ethylene-1,2-diamine) (55 mg, 0.38 mmol).

16c or **16d** were synthesized as describe above from **14c** (56 mg, 0.1 mmol) or **14b** (53 mg, 0.1 mmol), respectively, and diamine **15c**.

16e or **16f** were synthesized as described above from **14c** (56 mg, 0.1 mmol) or **14b**, respectively, (53 mg, 0.1 mmol) and diamine **15d**.

16i was synthesized as described above from **14d** (350 mg, 0.7 mmol) and **15b** (N,N,N'-triethyl-ethylene-1,2-diamine) (180 μ l, 1 mmol).

5

16a: Yield 120 mg (69%) as TFA salt

MS $[M+Na]^+ = 692.4$ (MW+Na calculated = 692.9 g/mol)

16b: Yield 48 mg (40%) as TFA salt

MS $[M+Na]^+ = 736.3$ (MW+Na calculated = 736.4 g/mol)

10

16c: Yield 8 mg (10%) as TFA salt

MS $[M+Na]^+ = 736.4$ (MW+Na calculated = 736.4 g/mol)

16d: Yield 20 mg (25%) as TFA salt

MS $[M+Na]^+ = 706.3$ (MW+Na calculated = 706.3 g/mol)

16e: Yield 2 mg (3%) as TFA salt

15

MS $[M+Na]^+ = 764.6$ (MW+Na calculated = 764.4 g/mol)

16f: Yield 6 mg (8%) as TFA salt

MS $[M+Na]^+ = 734.4$ (MW+Na calculated = 734.3 g/mol)

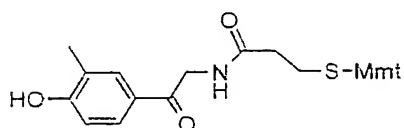
16i: Yield 152 mg (28%) as TFA salt

MS $[M+Na]^+ = 690.5$ (MW+Na calculated = 690.9 g/mol)

20

Synthesis of compound 17

25



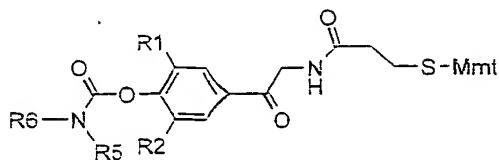
17

Amine **12b** (TFA salt) was coupled to compound **2** as described for compound **3a**.

30

17: Yield 608 mg (74%)

MS $[M+Na]^+ = 548.3$ (MW+Na calculated = 548.7 g/mol)

Synthesis of compounds 18a and 18b

18a R1 = R5 = Me, R2 = H, R6 = 3-(dimethylamino)propyl

18b R1 = R5 = Me, R2 = H, R6 = 2-(N-ethyl-N-methylamino)ethyl

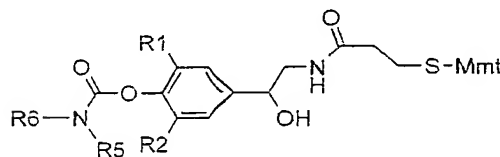
383 mg (0.729 mmol) of **17** were reacted with p-nitrophenyl-chloroformate and N,N,N'-trimethyl-1-propane-1,3-diamine (**15e**) or **15f**, respectively, to yield **18a** or **18b** as described for compound **16a**.

18a: Yield 287 mg (50%) as TFA salt

MS $[M+Na]^+ = 690.7$ (MW+Na calculated = 690,9 g/mol)

18b: Yield 148 mg (26%) as TFA salt

MS $[M+Na]^+ = 690.9$ (MW+Na calculated = 690,9 g/mol)

Synthesis of compounds 16g and 16h

16g R1 = R5 = Me, R2 = H, R6 = 3-(dimethylamino)propyl

16h R1 = R5 = Me, R2 = H, R6 = 2-(N-ethyl-N-methylamino)ethyl

18a (287 mg, 0.367 mmol, TFA salt) was dissolved in 5 ml methanol, NaBH₄ (41 mg, 1.07 mmol) was added and the mixture was stirred for 30 min at RT. 0.5 ml of acetic acid were added and **16g** was purified by RP-HPLC.

18b (8 mg, 0.010 mmol, TFA salt) was reacted as described above to yield **16h**.

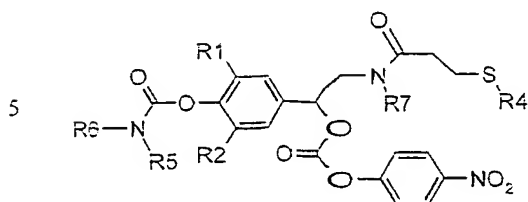
16g: Yield 201 mg (70%) as TFA-salt

MS $[M+Na]^+ = 692.7$ (MW+Na calculated = 692.9 g/mol)

16h: Yield 6 mg (77%) as TFA-salt

MS $[M+Na]^+ = 692.7$ (MW+Na calculated = 692.9 g/mol)

Synthesis of compounds 19a to 19i



19a R1 = R2 = R5 = Me, R6 = 2-(dimethylamino)ethyl, R7 = H, R4 = Mmt

19b R1 = OMe, R2 = H, R5 = Et, R6 = 2-(diethylamino)ethyl, R7 = H, R4 = Mmt

10 19c R1 = OMe, R2 = H, R5 = Me, R6 = 3-(N-ethyl-N-methylamino)propyl, R7 = Me, R4 = Mmt

19d R1 = R2 = H, R5 = Me, R6 = 3-(N-ethyl-N-methylamino)propyl, R7 = Me, R4 = Mmt

19e R1 = OMe, R2 = H, R5 = Et, R6 = 3-(diethylamino)propyl, R7 = Me, R4 = Mmt

15 19f R1 = R2 = H, R5 = Et, R6 = 3-(diethylamino)propyl, R7 = Me, R4 = Mmt

19g R1 = R5 = Me, R2 = H, R6 = 3-(dimethylamino)propyl, R7 = H, R4 = Mmt

19h R1 = R5 = Me, R2 = H, R6 = 2-(N-ethyl-N-methylamino)ethyl, R7 = H, R4 = Mmt

19i R1 = R2 = H, R5 = Et, R6 = 2-(diethylamino)ethyl, R7 = Me, R4 = Trt

20

Carbonates 19a to 19i were synthesized from 16a to 16i, respectively, as described for compound 7a.

19a: Yield 98 mg (72%) as TFA-salt

25 MS $[M+Na]^+ = 857.8$ (MW+Na calculated = 858.0 g/mol)

19b: Yield 6 mg (11%) as TFA-salt

MS $[M+Na]^+ = 901.8$ (MW+Na calculated = 901.5 g/mol)

19c: Yield 1 mg (15%) as TFA-salt

MS $[M+Na]^+ = 901.4$ (MW+Na calculated = 901.5 g/mol)

30 19d: Yield 8 mg (29%) as TFA-salt

MS $[M+Na]^+ = 871.4$ (MW+Na calculated = 871.4 g/mol)

19e: Yield 0.3 mg (18%) as TFA-salt

MS $[M+Na]^+ = 929.4$ (MW+Na calculated = 929.5 g/mol)

19f: Yield 4 mg (45%) as TFA-salt

MS $[M+Na]^+ = 899.7$ (MW+Na calculated = 899.6 g/mol)

19g: Yield 6 mg (6%) as TFA-salt

MS $[M+Na]^+ = 857.8$ (MW+Na calculated = 858.0 g/mol)

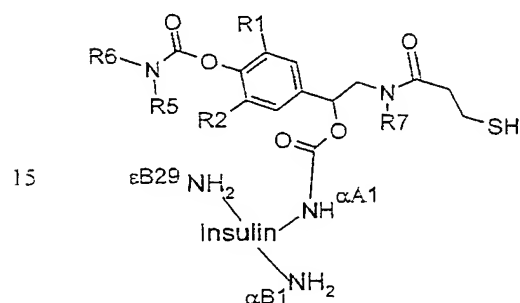
19h: Yield 0.8 mg (11%) as TFA-salt

5 MS $[M+Na]^+ = 857.7$ (MW+Na calculated = 858.0 g/mol)

19i: Yield 77 mg (49%) as TFA-salt

MS $[M+Na]^+ = 856.2$ (MW+Na calculated = 856.0 g/mol)

10 Synthesis of compounds **20a** to **20f**



20a R1 = R2 = R5 = Me, R6 = 2-(dimethylamino)ethyl, R7 = H

20 **20b** R1 = OMe, R2 = H, R5 = Et, R6 = 2-(diethylamino)ethyl, R7 = H

20c R1 = OMe, R2 = H, R5 = Me, R6 = 3-(N-ethyl-N-methylamino)propyl, R7 = Me

20d R1 = R2 = H, R5 = Me, R6 = 3-(N-ethyl-N-methylamino)propyl, R7 = Me

20e R1 = R5 = Me, R2 = H, R6 = 3-(dimethylamino)propyl, R7 = H

20f R1 = R5 = Me, R2 = H, R6 = 2-(N-ethyl-N-methylamino)ethyl, R7 = H

25

Insulin derivatives **20a**, **20b**, **20c**, **20d**, **20e**, or **20f** were synthesized from **19a**, **19b**, **19c**, **19d**, **19g**, or **19h** respectively, as described for compound **8a**.

20a MS $[M+3H]^{3+} = 2077.3$ $[M+4H]^{4+} = 1559.2$ (MW calculated = 6231.3 g/mol)

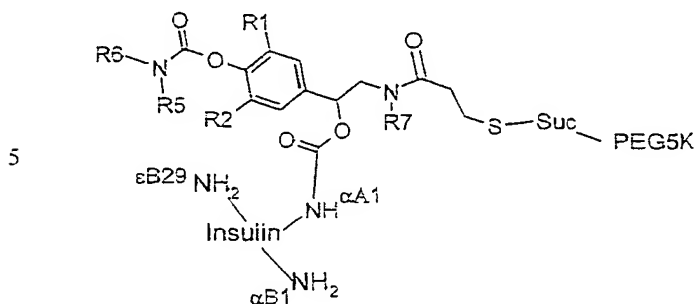
30 **20b** MS $[M+3H]^{3+} = 2093.0$ $[M+4H]^{4+} = 1569.6$ (MW calculated = 6274 g/mol)

20c MS $[M+3H]^{3+} = 2090.8$ $[M+4H]^{4+} = 1568.7$ (MW calculated = 6274 g/mol)

20d MS $[M+3H]^{3+} = 2081.3$ $[M+4H]^{4+} = 1561.8$ (MW calculated = 6244 g/mol)

20e MS $[M+3H]^{3+} = 2077.1$ $[M+4H]^{4+} = 1558.2$ (MW calculated = 6231.3 g/mol)

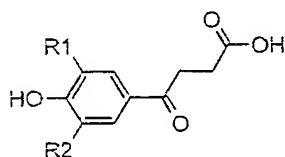
20f MS $[M+3H]^{3+} = 2076.7$ $[M+4H]^{4+} = 1559.3$ (MW calculated = 6231.3 g/mol)

Synthesis of compounds 21a to 21f (mono-pegylated insulin derivatives)

- 10 **21a** $R_1 = R_2 = R_5 = \text{Me}$, $R_6 = 2\text{-(dimethylamino)ethyl}$, $R_7 = \text{H}$
 21b $R_1 = \text{OMe}$, $R_2 = \text{H}$, $R_5 = \text{Et}$, $R_6 = 2\text{-(diethylamino)ethyl}$, $R_7 = \text{H}$
 21c $R_1 = \text{OMe}$, $R_2 = \text{H}$, $R_5 = \text{Me}$, $R_6 = 3\text{-(N-ethyl-N-methylamino)propyl}$, $R_7 = \text{Me}$
 21d $R_1 = R_2 = \text{H}$, $R_5 = \text{Me}$, $R_6 = 3\text{-(N-ethyl-N-methylamino)propyl}$, $R_7 = \text{Me}$
 21e $R_1 = R_5 = \text{Me}$, $R_2 = \text{H}$, $R_6 = 3\text{-(dimethylamino)propyl}$, $R_7 = \text{H}$
 15 **21f** $R_1 = R_5 = \text{Me}$, $R_2 = \text{H}$, $R_6 = 2\text{-(N-ethyl-N-methylamino)ethyl}$, $R_7 = \text{H}$

Insulin derivatives **21a**, **21b**, **21c**, **21d**, **21e**, or **21f** were synthesized from compound **20a**, **20b**, **20c**, **20d**, **20e**, or **20f**, respectively, as described for compound **9a**.

- 20 **21a** through **21f**: SEC retention time: 19.5 min

Synthesis of compounds 23a and 23b

23a R1 = Me, R2 = H

23b R1 = R2 = Me

o-Cresol (**22a**) (1 eq), succinic anhydride (1 eq), and AlCl_3 (3 eq) in nitrobenzene were heated to 100 °C for 1 h. The reaction mixture was poured on HCl/ice and extracted with ether. The organic layer was extracted with 1 N NaOH and the aqueous layer was acidified with concentrated HCl. The aqueous layer was extracted with ether and the ether was evaporated. **23a** was purified by RP-HPLC.

23b was synthesized from 2,6-dimethylphenol (**22b**) as described above.

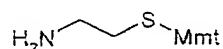
23a: Yield 552 mg (31%)

MS $[\text{M}+\text{Na}]^+ = 231.0$ (MW+Na calculated = 231.2 g/mol)

NMR (300 MHz, DMSO- d_6) δ [ppm] = 12.05 (bs, 1H, CO_2H), 10.23 (s, 1H, phenol OH), 7.74 (s, 1H, CH_{ar}), 7.7 (d, 1H, CH_{ar} , $^3J_{\text{H,H}} = 8.4$ Hz), 6.86 (d, 1H, CH_{ar} , $^3J_{\text{H,H}} = 8.4$ Hz), 3.13 (t, 2H, C(O)CH_2 , $^3J_{\text{H,H}} = 6.4$ Hz), 2.53 (t, 2H, CH_2CO_2 , $^3J_{\text{H,H}} = 6.4$ Hz), 2.16 (s, 3H, CH_3)

23b: Yield 166 mg (15%)

MS $[\text{M}+\text{Na}]^+ = 245.4$ (MW+Na calculated = 245.2 g/mol)

Synthesis of compound 24

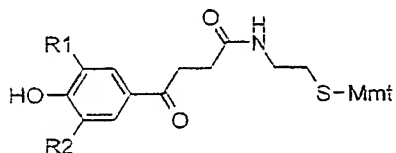
1.85 g (16.02 mmol) cysteamine hydrochloride were dissolved in 15 ml of TFA and 2.47 g (8.01 mmol) MmtCl were added. After stirring the mixture at RT for 20 min the solvent was evaporated in vacuo. The residue was dissolved in diethyl ether and extracted with saturated aqueous NaHCO_3 , 1N H_2SO_4 and brine. The solvent was evaporated and **24** was purified by RP-HPLC.

24: Yield 1.11 g (30%) as TFA salt

TLC (AcOEt/Et₃N 99/1), R_f = 0.24

Synthesis of compounds **25a** and **25b**

5



25a R1 = Me, R2 = H

10 **25b** R1 = R2 = Me

23a (1eq), HOBt (1.1 eq) and DIC (1 eq) were dissolved in DMF and stirred at RT for 30 min. **24** (TFA salt, 1 eq) and DIEA (3 eq) were added and the solution was stirred for 60 min. Acetic acid was added and **25a** was purified by RP-HPLC.

15 **25b** was synthesized from **23b** as described above.

25a: Yield 552 mg (25%)

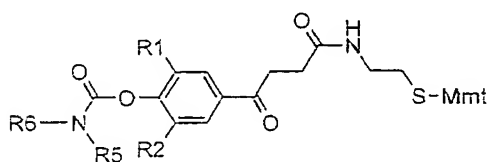
MS [M+Na]⁺ = 562.7 (MW+Na calculated = 562.7 g/mol)

25b: Yield 15 mg (40%)

20 MS [M+Na]⁺ = 576.6 (MW+Na calculated = 576.6 g/mol)

Synthesis of compounds **26a** and **26b**

25



26a R1 = Me, R2 = H, R5 = R6 = 3-(dimethylamino)propyl

26b R1 = R2 = R5 = Me, R6 = 2-(dimethylamino)ethyl

30 267 mg (0.495 mmol) **25a** was reacted with p-nitrophenylchloroformate and N-(3-dimethylamino-propyl)-N',N'-dimethyl-propane-1,3-diamine (**15g**) to yield **26a** as described for compound **16a**.

26b was synthesized as described above using 15 mg **25b** and N,N,N'-trimethyl-ethane-1,2-diamine (**15a**).

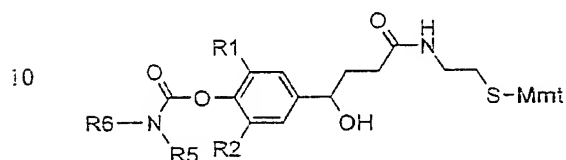
26a: Yield 282 mg (58%) as double TFA salt

MS $[M+Na]^+ = 775.2$ (MW+Na calculated = 776.0 g/mol)

26b: Yield 17 mg (70%) as TFA salt

5 MS $[M+Na]^+ = 704.5$ (MW+Na calculated = 704.6 g/mol)

Synthesis of compounds **27a** and **27b**



27a R1 = Me, R2 = H, R5 = R6 = 3-(dimethylamino)propyl

27b R1 = R2 = R5 = Me, R6 = 2-(dimethylamino)ethyl

15

26a (272 mg, 0.277 mmol, double TFA salt) was dissolved in 5 ml methanol, NaBH₄ (42 mg, 1.09 mmol) was added and the mixture was stirred for 30 min at RT. 0.5 ml of acetic acid were added and **27a** was purified by RP-HPLC.

Alcohol **27b** was synthesized likewise from **26b** (17 mg, 25 μmol, TFA salt).

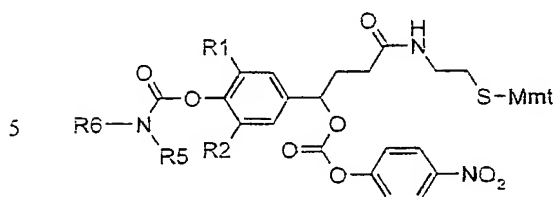
20

27a: Yield 142 mg (52%) as double TFA salt

MS $[M+Na]^+ = 777.9$ (MW+Na calculated = 778.0 g/mol)

27b: Yield 6 mg (40%) as TFA salt

MS $[M+Na]^+ = 706.5$ (MW+Na calculated = 706.6 g/mol)

Synthesis of compounds 28a and 28b

28a R1 = Me, R2 = H, R5 = R6 = 3-(dimethylamino)propyl

28b R1 = R2 = R5 = Me, R6 = 2-(dimethylamino)ethyl

10

Carbonates **28a** or **28b** were synthesized from **27a** or **27b**, respectively, as described for compound **7a**.

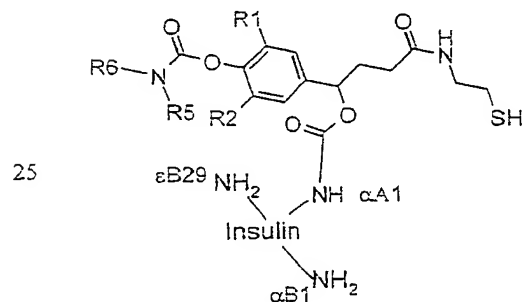
15 **28a**: Yield 1 mg (29%)

MS $[M+Na]^+ = 942.9$ (MW+Na calculated = 943.2 g/mol)

28b: Yield 1.5 mg (19%)

MS $[M+Na]^+ = 871.6$ (MW+Na calculated = 871.7 g/mol)

20 Synthesis of compounds 29a and 29b



29a R1 = Me, R2 = H, R5 = R6 = 3-(dimethylamino)propyl

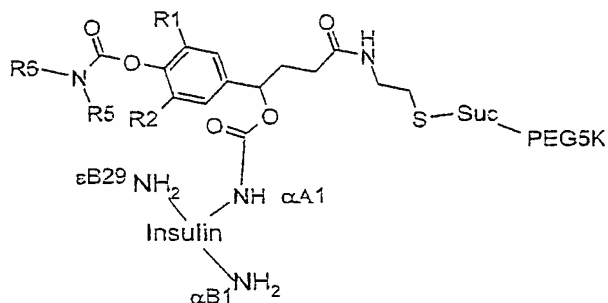
30 **29b** R1 = R2 = R5 = Me, R6 = 2-(dimethylamino)ethyl

Insulin derivatives **29a** or **29b** were synthesized from **28a** or **28b**, respectively, as described for compound **8a**.

29a MS $[M+3H]^{3+} = 2105.8$ $[M+4H]^{4+} = 1580.2$ (MW calculated = 6316.4 g/mol)

29b MS $[M+3H]^{3+} = 2081.8$ $[M+4H]^{4+} = 1562.4$ (MW calculated = 6244 g/mol)

Synthesis of mono-pegylated insulin derivatives **30a** and **30b**



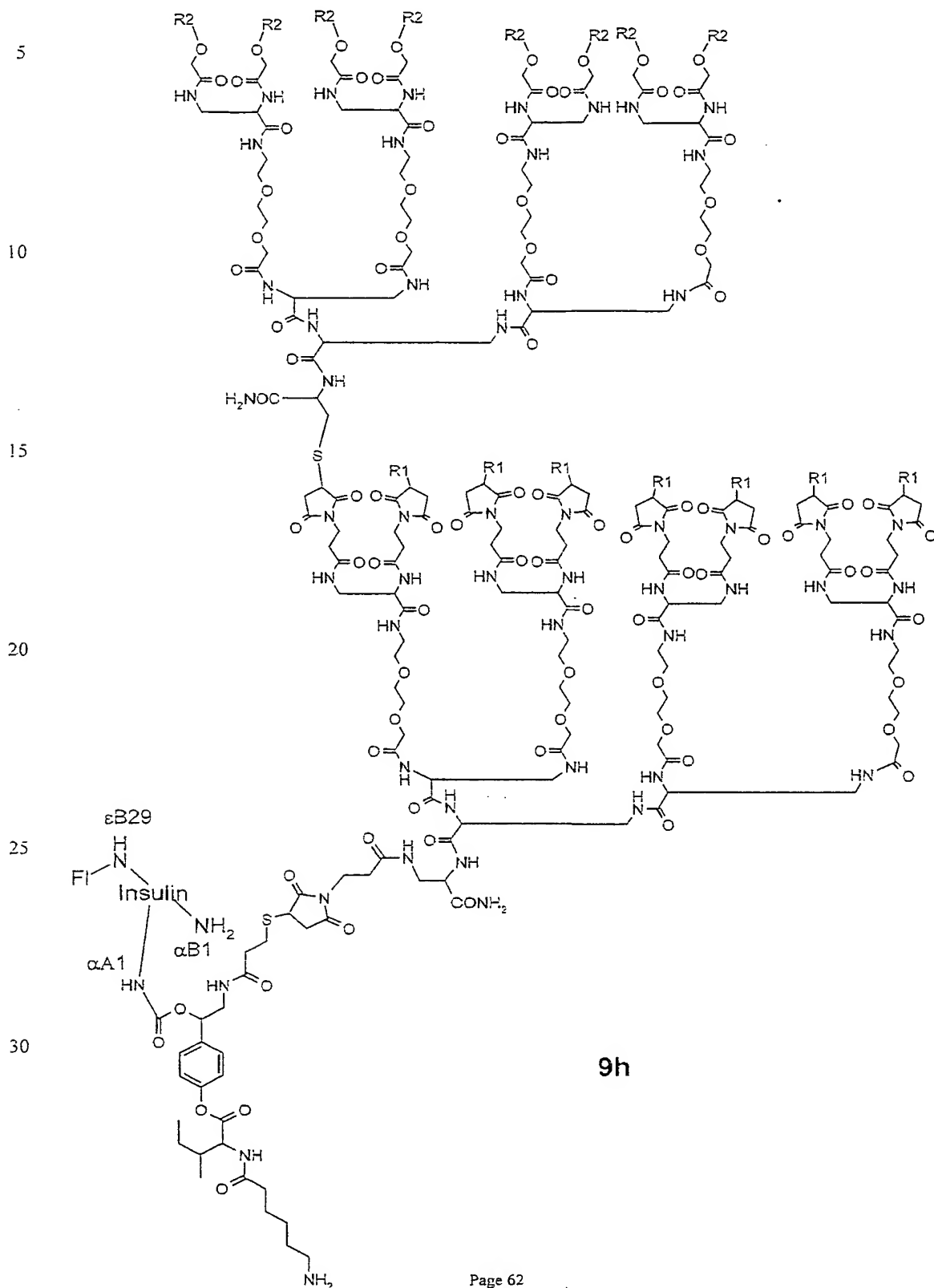
30a R1 = Me, R2 = H, R5 = R6 = 3-(dimethylamino)propyl

30b R1 = R2 = R5 = Me, R6 = 2-(dimethylamino)ethyl

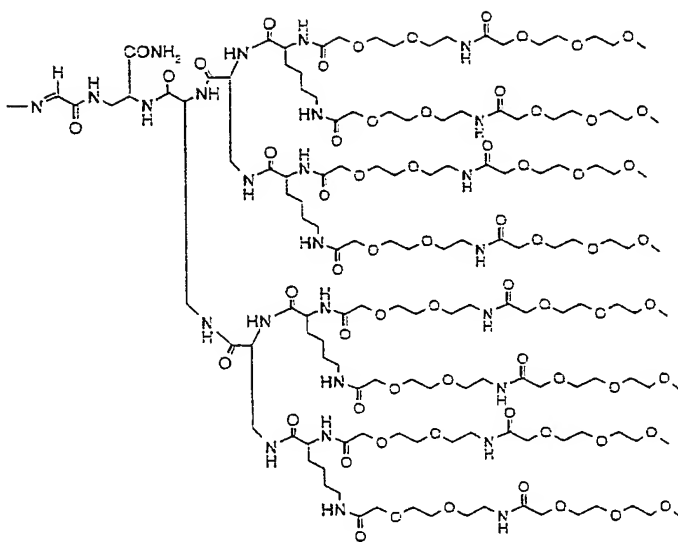
Insulin derivatives **30a** or **30b** were synthesized from **29a** or **29b**, respectively, as described for compound **9a**.

30a and **30b**: SEC retention time: 19.5 min

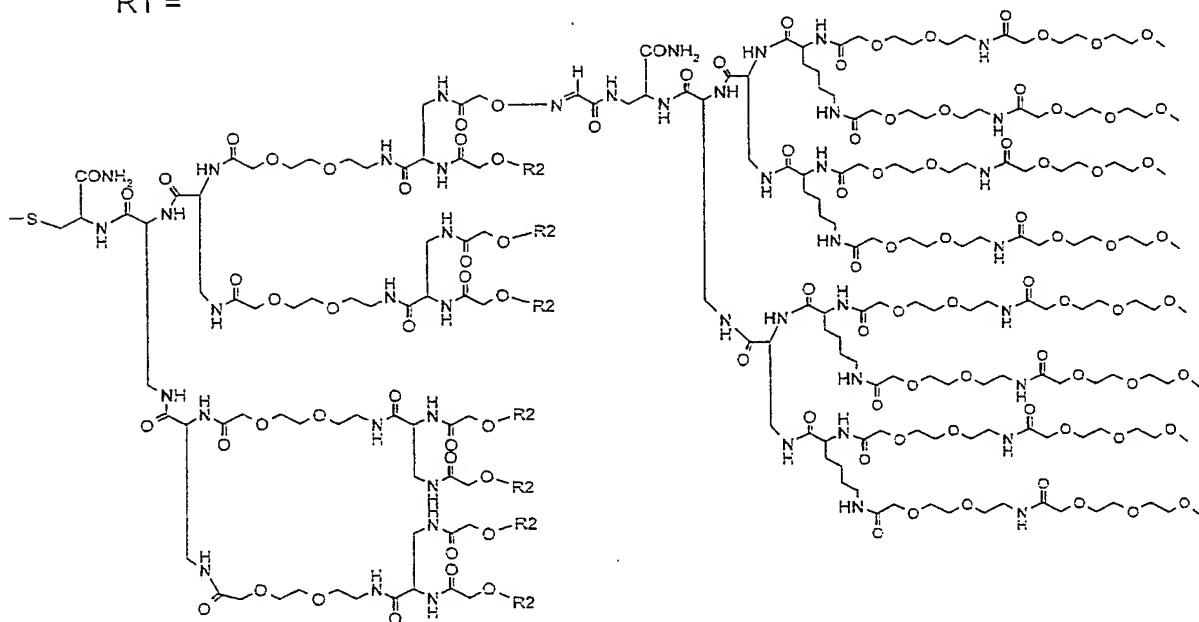
Synthesis of a polymeric prodrug according to formula Ia with an ester-linked
masking group and a dendritic carrier (9h)



R2 =



R1 =



S.

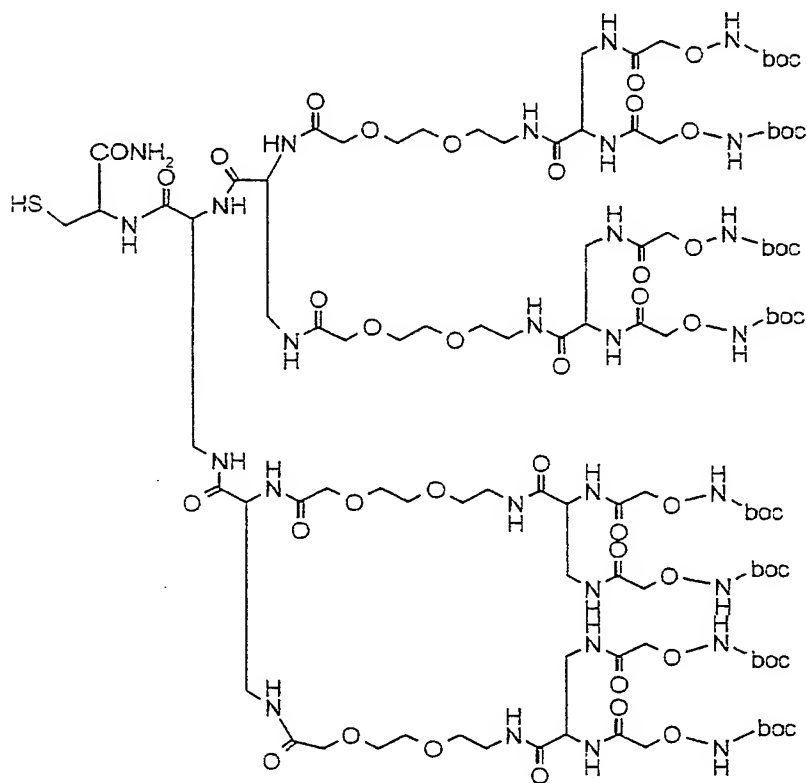
31

5

10

MS: $[M+H]^+ = 2494.6$ (MW calculated = 2495.4 g/mol)

Synthesis of compound **32**



32

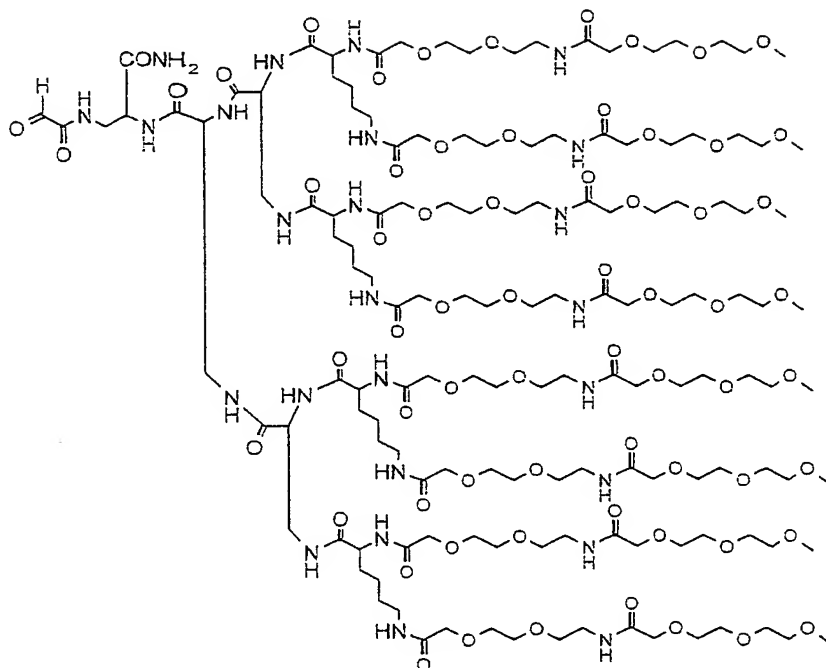
Compound **32** was obtained according to the standard solid-phase synthesis protocol.

- 5 The amino acids Fmoc-Cys(Mmt)-OH, Fmoc-Dpr(Fmoc)-OH, Fmoc-Dpr(Fmoc)-OH, Fmoc-Ado-OH, and Fmoc-Dpr(Fmoc)-OH were coupled to NovaSynTG Sieber amide resin.

- After final fmoc removal the resin was agitated with 3 eq Boc-aminoxycetic acid, 3 eq DIC, and 3 eq HOBt in relation to amino groups in DMF for 30 min. **32** was
 10 cleaved from resin with DCM/TFA/TES 97/1/2 (v/v/v). After addition of 0.8 eq pyridine in relation to TFA the solvent was evaporated and product **32** was purified by RP-HPLC.

MS: $[M+H]^+ = 2688.2$ g/mol (MW calculated = 2688.8 g/mol)

Synthesis of compound 33

**33**

5

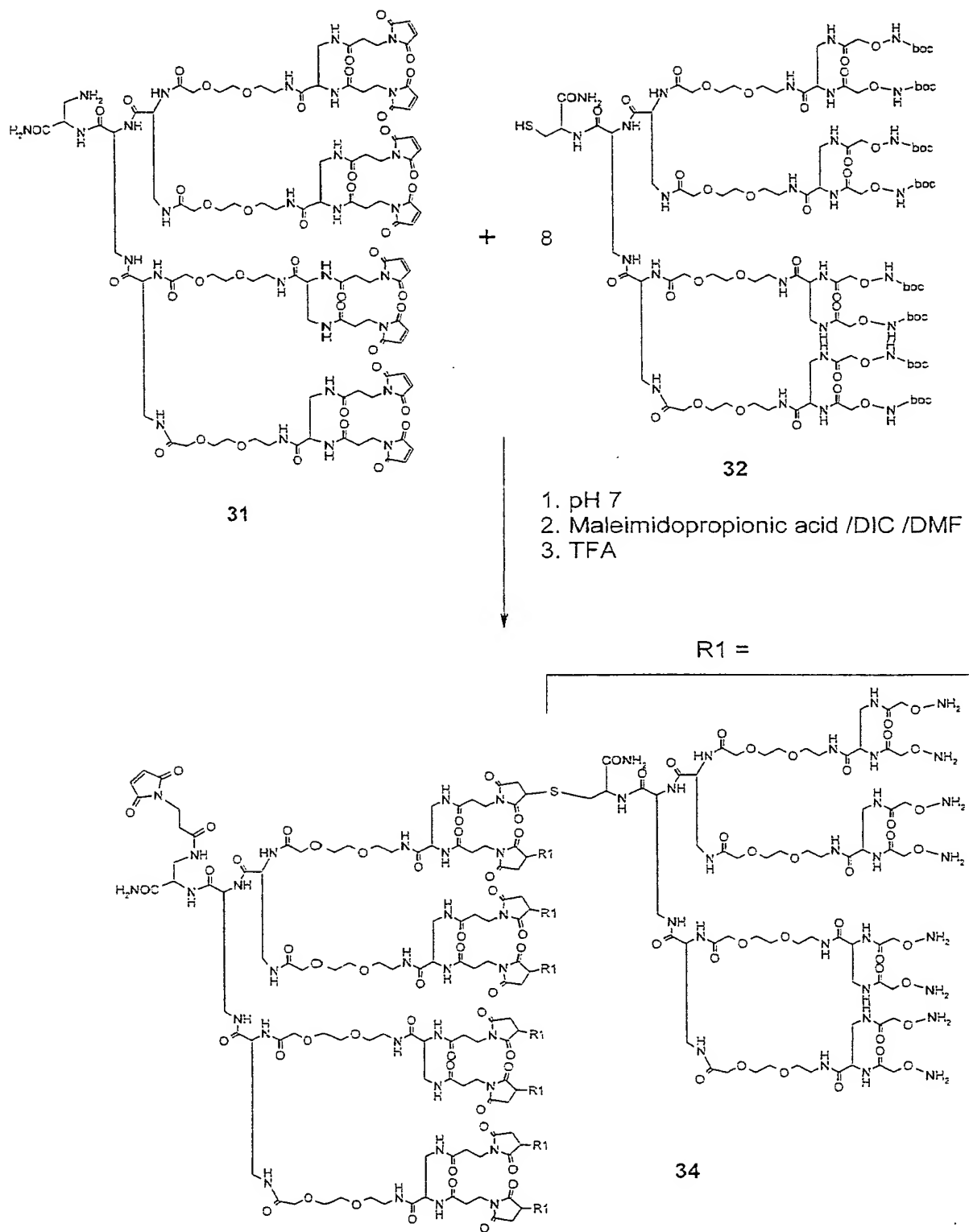
Compound 33 was obtained according to the standard solid-phase synthesis protocol. The amino acids Fmoc-Dpr(ivDde)-OH, Fmoc-Dpr(Fmoc)-OH, Fmoc-Dpr(Fmoc)-OH, Fmoc-Lys(Fmoc)-OH, and Fmoc-Ado-OH were coupled to NovaSyn TG Sieber amide resin.

- 10 After final fmoc removal the resin was agitated with 3 eq 3,6,9-trioxadecanoic acid, 3 eq PyBOP, and 6 eq DIEA in relation to amino groups in DMF for 60 min.

To cleave the ivDde protecting group, the resin was treated three times with 2% hydrazine in DMF. After washing, 3 eq Fmoc-Ser-OH was coupled with 3eq DIC and 3 eq HOBt for 30 min. After final fmoc removal resin was washed and the product
 15 was cleaved from resin with DCM/TFA/TES 88/10/2 (v/v/v). Solvent was evaporated and the residue was oxidized with 10 eq sodium periodate in 3/2 (v/v) 0.1 M sodium phosphate pH 7/acetonitrile for 15 min to yield 33. Product 33 was purified by RP-HPLC and lyophilized.

MS: $[M+H]^+ = 3372.1$ g/mol (3372.8 g/mol)

Synthesis of compound 34



6 mg (2.4 μ mol) of compound **31** were dissolved in 1 ml 2/1 (v/v) acetonitrile/0.1 M sodium phosphate buffer pH 7 and 65 mg (24.2 μ mol) of compound **32** were added.

5 The solution was stirred at room temperature for 2 h and then the product purified by RP-HPLC and lyophilized (yield: 45 mg (78%)).

The lyophilized product (45 mg) was dissolved in 0.5 ml DMF and 10 μ l DIEA were added. 5 mg (30 μ mol) 3-maleimidopropionic acid and 4.7 μ l (30 μ mol) DIC in 150 μ l
10 DMF were added and the reaction mixture was stirred at room temperature for 20 min, the product purified by RP-HPLC and lyophilized.

The lyophilized product was incubated for 10 min in 95/5 (v/v) TFA/water and then the solvent was removed in a stream of nitrogen. Product **34** was purified by RP-
15 HPLC and lyophilized (overall yield for all three steps: 20 mg (47%)).

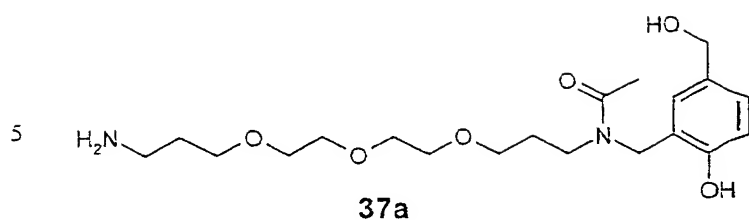
MS: 17700 – 18200 (broad peak) (MW calculated = 17749 g/mol)

Synthesis of compound **9h**

20 1.5 mg (225 nmol) **8g** and 5 mg (280 nmol) **34** were mixed, dissolved in 300 μ l 2/1 (v/v) 0.1 M sodium phosphate buffer pH 7/acetonitrile and incubated for 15 min at room temperature. The product was purified by RP-HPLC and lyophilized. (yield 4 mg, 160 nmol, 70%)

25 The lyophilized product was dissolved in 200 μ l 0.1 M sodium citrate buffer pH 1.5 and 69 mg (20.5 μ mol) **33** in 200 μ l 2/1 (v/v) acetonitrile/sodium citrate buffer pH 1.5 were added. The mixture was stirred at room temperature for 24 h and product **9h** was purified by size exclusion chromatography (column: Superdex 200, buffer: 10 mM
30 HEPES pH 7.4, 0.005% Tween-20, 3mM EDTA, flow rate: 0.75 ml/min)

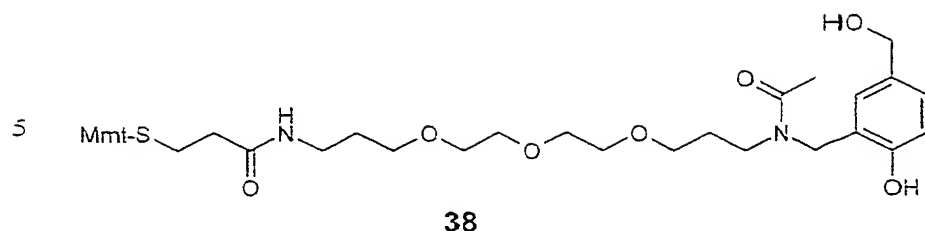
SEC elution time: 15 min

Synthesis of compound 37a

250 mg (0.35 mmol) 2-chlorotrityl chloride resin (loading 1.4 mmol/g) was incubated
 10 for 1.5 h with 308 mg (4 eq., 1.4 mmol) 4,7,10-trioxatridecane-1,13-diamine in 4 ml
 DCM to yield **35a**. The resin was washed with DCM and dried. 107 mg (0.7 mmol)
 HOBt, 110 μ l (0.7 mmol) DIC, and 150 mg (0.9 mmol) 5-formyl salicylic acid in 3 ml
 DMF were added and the resulting suspension was stirred for 1 h at RT to yield **36a**.
 After washing with DCM and THF, the resin was suspended in 6 ml THF and 3 ml (3
 15 mmol) BH_3 THF (1 M in THF, 8.5 eq.) were added dropwise. The reaction mixture
 was stirred for 18 h at 45°C under nitrogen atmosphere. After cooling 4 ml THF, 0.8
 ml DIEA and 1.6 ml MeOH were added successively. 210 mg (0.84 mmol) I_2 (as a
 concentrated THF solution) were added and the suspension was stirred for 1 h. The
 resin was repeatedly washed (three times each) with THF, DMF, MeOH, and DCM.
 20 The dried resin was reacted with 107 mg (0.7 mmol) HOBt, 110 μ l (0.7 mmol) DIC,
 and 55 μ L (0.9 mmol) AcOH in 3 ml DMF for 1h. After washing of the resin with
 DMF and DCM compound **37a** was cleaved from resin with 2/1 (v/v) HFIP/DCM
 (two times for 30 min). The volatile components were evaporated and the product **37a**
 was used in the following step without further purification.

25 **37a**: Yield 29 mg (20%) as TFA salt

MS $[\text{M}+\text{Na}]^+ = 421.4$ (MW+Na calculated = 421.5 g/mol)

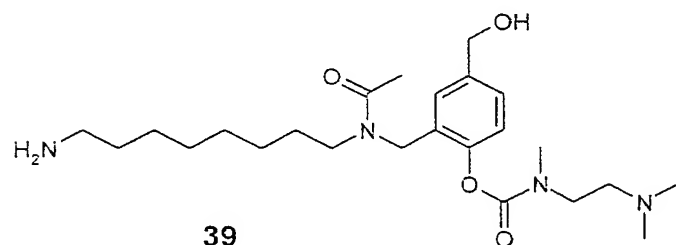
Synthesis of compound 38

24 mg (0.06 mmol) **37a**, 31 mg (0.06 mmol) PyBOP, 32 μ l (0.18 mmol) DIEA, and
 10 23 mg (0.06 mmol) **2** in 0.5 ml DMF were reacted for 50 min at room temperature.
 After addition of 50 μ l acetic acid product **38** was purified by RP-HPLC.

38: Yield 7 mg (15%)

MS $[M+Na]^+ = 781.3$ (MW+Na calculated = 781.6 g/mol)

15

Synthesis of compound 39

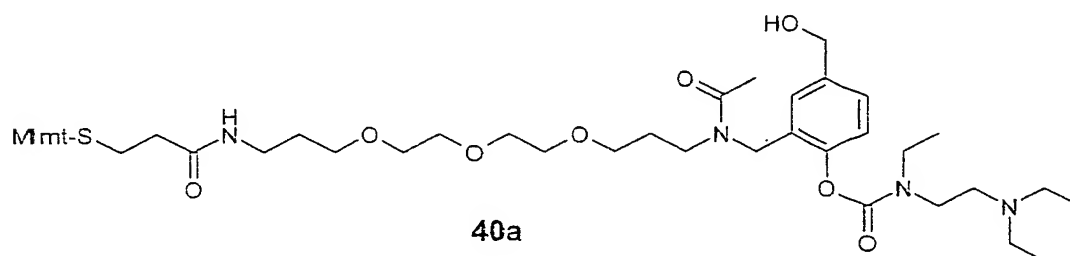
20 300 mg (0.42 mmol) 2-chlorotriyl chloride resin (loading 1.4 mmol/g) was incubated
 for 1.5 h with 245 mg (4 eq., 1.7 mmol) 1,8-diaminooctane in 4 ml DCM to yield **35b**.
 The resin was washed with DCM and dried. 107 mg (0.7 mmol) HOBt, 110 μ l (0.7
 mmol) DIC, and 150 mg (0.9 mmol) 5-formyl salicylic acid in 3 ml DMF were added
 and the resulting suspension was stirred for 1 h at RT to yield **36b**. After washing with
 25 DCM and THF, the resin was suspended in 6 ml THF and 3 ml (3 mmol) BH_3 THF (1
 M in THF) were added dropwise. The reaction mixture was stirred for 18 h at 45°C
 under nitrogen atmosphere. After cooling 4 ml THF, 0.8 ml DIEA and 1.6 ml MeOH
 were added successively. 210 mg (0.84 mmol) I_2 (as a concentrated THF solution)
 were added and the suspension was stirred for 1 h. The resin was repeatedly washed

(three times each) with THF, DMF, MeOH, and DCM. The dried resin was reacted with 107 mg (0.7 mmol) HOBt, 110 μ l (0.7 mmol) DIC, and 55 μ L (0.9 mmol) AcOH in 3 ml DMF for 1h. After washing of the resin with DMF and DCM compound **37b**, 78 mg (0.39 mmol) p-nitrophenylchloroformate, and 210 μ l (1.2 mmol) DIEA in 1/1 (v/v) THF/DCM were reacted for 30 min at RT. The separated resin was suspended in 1/1 (v/v) THF/DCM and 210 μ l (1.2 mmol) N,N,N'-trimethylethylenediamine were added. The resulting suspension was stirred for 25 min at RT. The resin was separated and washed with DCM. Product **39** was cleaved from resin with 2/1 (v/v) HFIP/DCM (two times for 30 min). The volatile components were evaporated and the product **39** was purified by HPLC.

39: Yield 16 mg (8%) as TFA salt

MS $[M+Na]^+ = 473.5$ (MW+Na calculated = 473.3 g/mol)

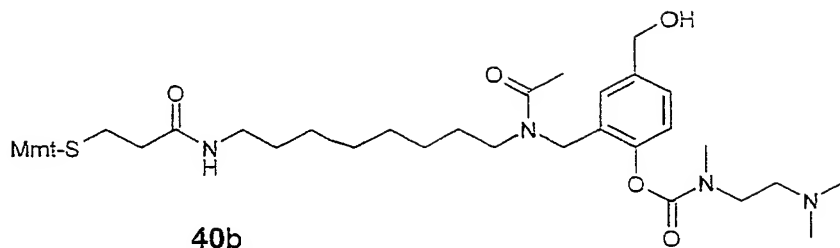
Synthesis of compound **40a**



38 (7 mg, 9 μ mol) was dissolved in 200 μ l of dry THF. p-Nitrophenylchloroformate (2.0 mg, 10 μ mol) and DIEA (4.4 μ l, 25 μ mol) were added and the mixture was stirred for 30 min at RT. N,N,N'-Triethylethylenediamine (**15b**) (18 μ l, 0.1 mmol) was added and stirring was continued for 30 min. The solvent was removed *in vacuo*, 10 μ l of AcOH were added and **40a** was purified by RP-HPLC.

40a: Yield 1 mg (11%) as TFA salt

MS $[M+Na]^+ = 951.1$ (MW+Na calculated = 951.8 g/mol)

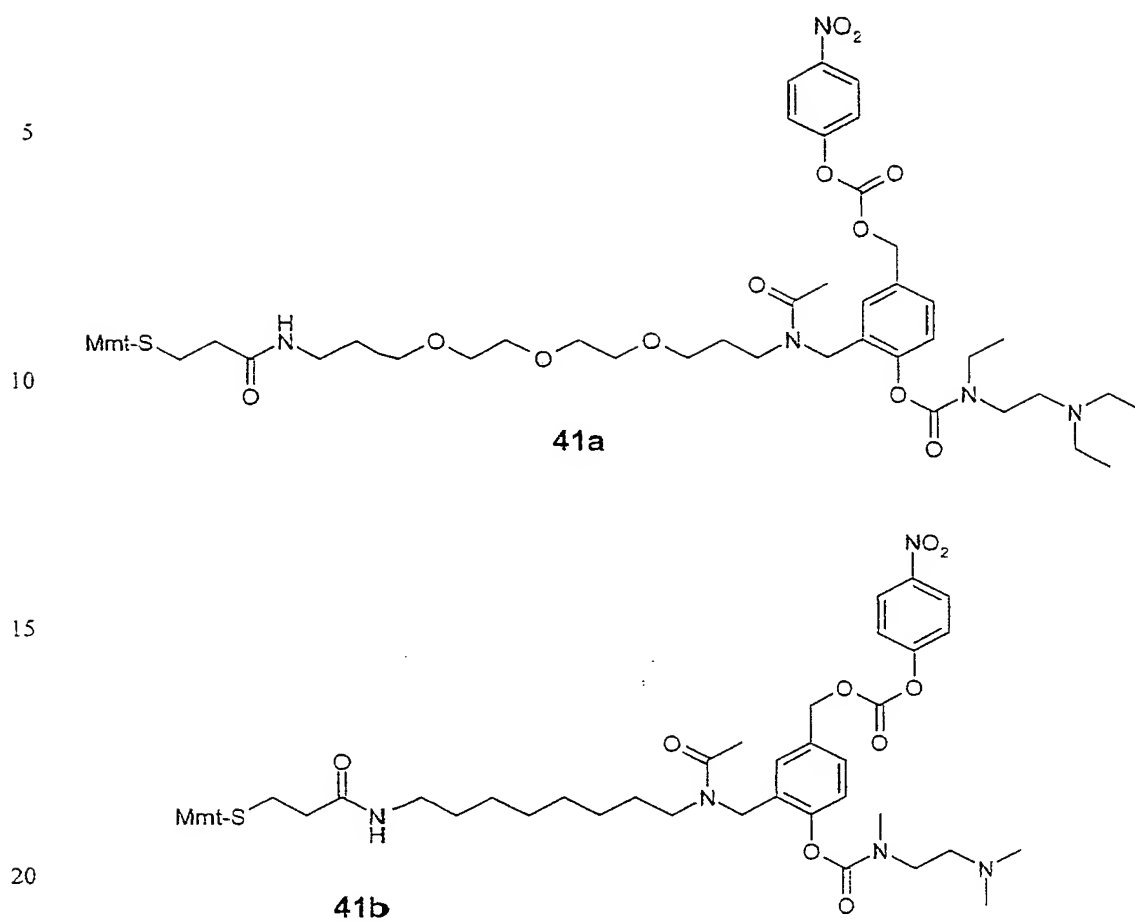
Synthesis of compound 40b

5

15 mg (33 μ mol) **39**, 18 mg (33 μ mol) PyBOP, 23 μ l (0.13 mmol) DIEA, and 13 mg (35 μ mol) **2** in 0.5 ml DMF were reacted for 45 min at room temperature. After addition of 50 μ l acetic acid product **40** was purified by RP-HPLC.

10 **40b**: Yield 10 mg (37%) as TFA salt

MS $[M+H]^+ = 811.5$ (MW+Na calculated = 810.5 g/mol)

Synthesis of compound 41a and 41b

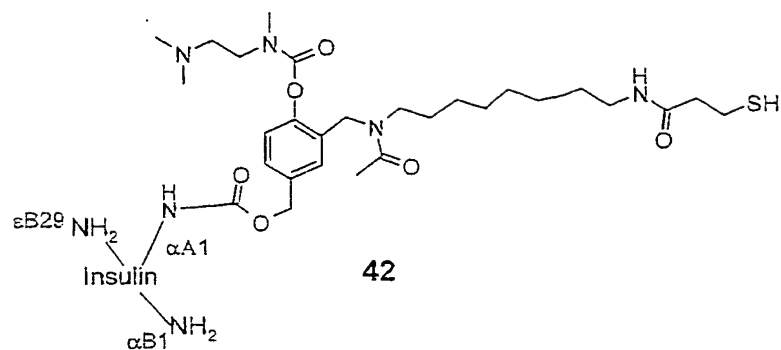
Carbonate 41a or 41b was synthesized from 40a or 40b as described for compound 7a

41a: Yield 0.4 mg as TFA salt

MS $[M+Na]^+ = 1116.8$ (MW+Na calculated = 1116.9 g/mol)

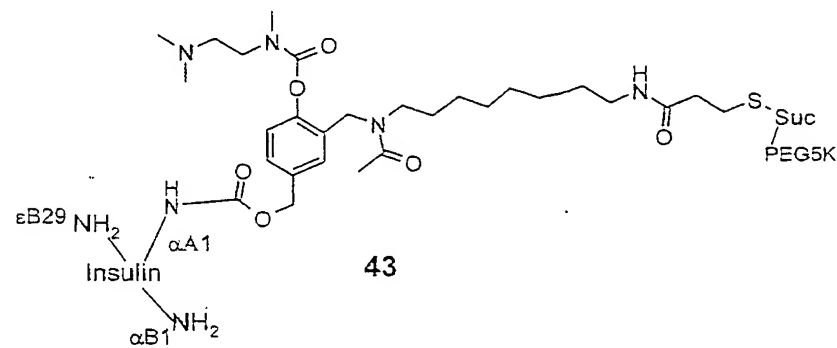
41b: Yield 2 mg (16%) as TFA salt

MS $[M+H]^+ = 976.8$ (MW calculated = 975.8 g/mol)

Synthesis of compound 42

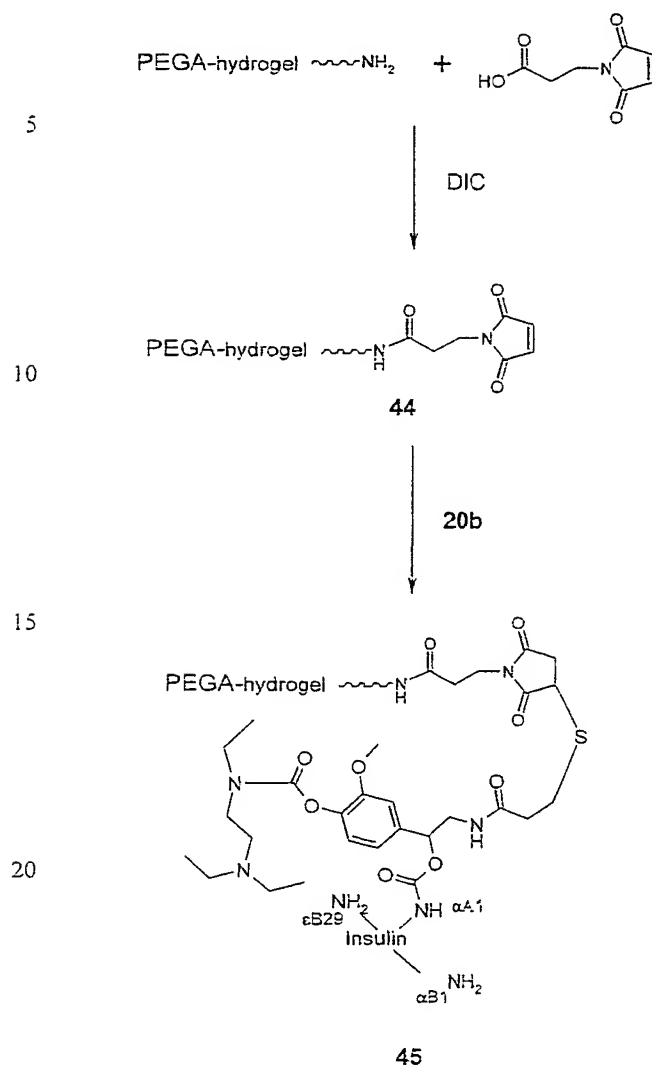
Insulin derivative **42** was synthesized from **41b** as described for compound **8a**.

42 MS $[M+3H]^{3+} = 2124.5$ $[M+4H]^{4+} = 1594.6$ (MW calculated = 6371 g/mol)

Synthesis of compound 43

Insulin derivative **43** was synthesized from **42** as described for compound **9a**.

43: SEC retention time: 18.0 min

Synthesis of rh-insulin loaded PEGA hydrogel 45

Maleimide derivatization of polyacrylamide based hydrogel (PEGA):

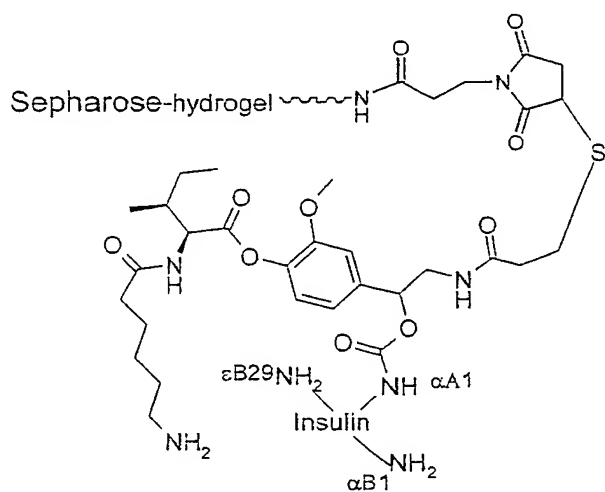
NH₂-PEGA hydrogel beads with 0.4 mmol/g loading and 150-300 μm bead size were purchased from Novabiochem.

2.5 g methanol-wet NH₂-PEGA-hydrogel (0.4 mmol/g NH₂-loading) was weighed into a syringe equipped with a polypropylene frit. Maleimide loading was adjusted by acylation employing a mixture of activated maleimidopropionic acid and acetic acid as described in the following. The hydrogel was washed 5 times with DMF and reacted with 13.5 mg (0.08 mmol) 3-maleimidopropionic acid, 115.2 μl (1.92 mmol)

acetic acid and 313 μ l (2 mmol) DIC in 4 ml DMF for 30 min. The maleimide derivatized hydrogel **44** was washed 10 times with DMF and DCM and finally with acetonitrile.

30 mg of maleimide derivatized resin **44** (loading 16 μ mol/g) was reacted with 3 mg of compound **20b** (480 nmol, 1.06 eq) in 600 μ l 20/80 (v/v) acetonitrile/50 mM phosphate buffer (pH 7.4) for 10 min to give rh-insulin loaded hydrogel **45**. The hydrogel **45** was washed 5 times with 50/50 (v/v) acetonitrile/water and three times with acetonitrile and dried under vacuum.

Synthesis of rh-insulin carbohydrate-based hydrogel **46**



46

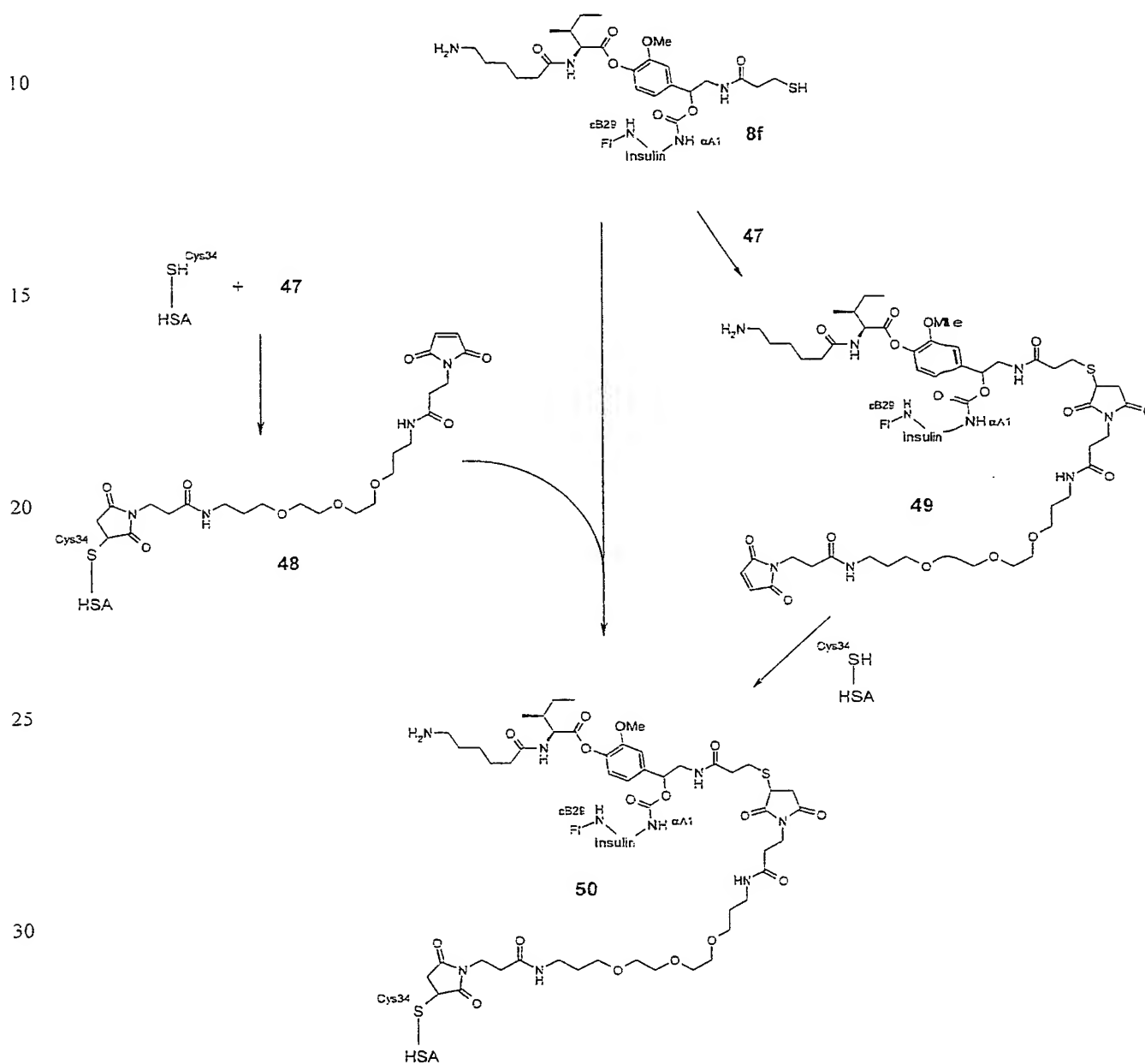
NHS-activated "Sepharose 4 Fast Flow" hydrogel beads (chemically crosslinked agarose, crosslinker epichlorhydrin) were purchased from Amersham.

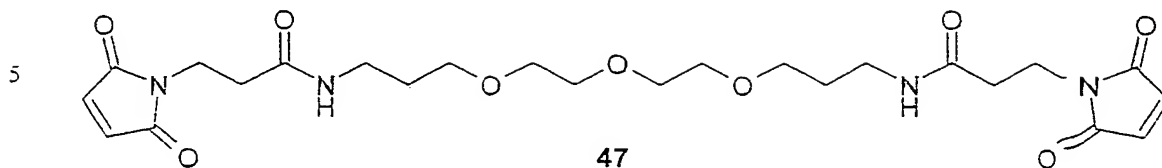
1.5 g ethanol-wet Sepharose hydrogel (150 mg dry hydrogel) was weighed into a syringe equipped with a polypropylene frit and reacted with 1 M 4,7,10-trioxatridecane-1,13-diamine in DMF for 30 min. After 5 washing steps with DMF, hydrogel was reacted with 8.5 mg (0.05 mmol) 3-maleimidopropionic acid, 57 μ l (0.95 mmol) acetic acid, 151 mg (1 mmol) HOBt and 158 μ l (1 mmol) DIC in 4 ml DMF for 30 min to give maleimide derivatized hydrogel. The hydrogel was washed 10 times with DMF and finally with acetonitrile.

1.5 mg **8c** was dissolved in 25/75 (v/v) acetonitrile/50 mM phosphate buffer pH 7.4 and reacted with 10.8 mg maleimide derivatized hydrogel for 10 min. The rh-insulin loaded hydrogel **46** was washed five times with 50/50 (v/v) acetonitrile/water and three times with acetonitrile and dried under vacuum.

5

Synthesis scheme of Fluorescein-Insulin-rHSA (**50**)



Synthesis of bismaleimide 47

3-Maleimido-propionic acid (92 mg, 0.54 mmol) in 200 μ l DMF was reacted with
 DIC (78 μ l, 0.50 mmol) at RT for 15 min. 4,7,10-Trioxa-tridecan-1,13-diamine (43.5
 10 μ l, 0.20 mmol) was added and the mixture was stirred for 30 min at RT.
 After addition of 800 μ l 1/4 (v/v) acetic acid/water 47 was purified by RP-HPLC.

47: Yield 23 mg (22%)

MS $[M+Na]^+ = 545.5$ (MW+Na calculated = 545.6 g/mol)

Synthesis of rHSA-maleimide (48)

66.5 μ l 3 mM rHSA solution in 145 mM NaCl, 32 mM sodium octanoate, 0.0015%
 Tween-80 was mixed with 66.5 μ l 0.5 M phosphate buffer pH 7.0. 0.41 mg
 20 bismaleimide 47 (0.8 μ mol) were added and the mixture was reacted for 15 min at
 RT. Compound 48 was purified by SEC (column: Superdex 200, flow rate: 0.75
 ml/min) using 10 mM HEPES buffer pH 7.4, 150 mM NaCl, 3 mM EDTA, and
 0.005% Tween as mobile phase. (Yield: 2.6 ml 77.5 μ M 48)

25 SEC retention time: 17.1 min (280 nm)

ESI-MS = 66988 (MW calculated = 66984 g/mol)

Synthesis of fluorescein-insulin-linker-maleimide (49)

30 40 μ l of 2.4 mM bismaleimide 47 in 1/1 (v/v) acetonitrile/water (96 nmol) were
 mixed with 40 μ l 0.5 M sodium borate buffer pH 5.8. 24 nmol 8f in 16.8 μ l 1/1 (v/v)
 acetonitrile/water were added and the mixture was incubated for 10 min at RT. 5 μ l
 AcOH were added and 49 was purified by RP-HPLC.

ESI-MS = 7211 (MW calculated = 7211 g/mol)

Synthesis of fluorescein-insulin-linker-rHSA 50

a) from 49 and rHSA

5 b) from 48 and 8f

a)

30 μ l of 80 μ M 49 in 1/1 (v/v) acetonitrile/water (2.4 nmol) were mixed with 70 μ l
0.25 M sodium phosphate buffer pH 6.4. 8 μ l 3 mM rHSA in 145 mM NaCl 32 mM
10 sodium octanoate, 0.0015% Tween-80, (24 nmol) was added and the mixture was
incubated at RT for 20 min.

Compound 50 was purified by SEC (column: Superdex 200, flow rate: 0.75 ml/min)
using 10 mM HEPES buffer pH 7.4, 150 mM NaCl, 3 mM EDTA, and 0.005%
Tween as mobile phase.

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SEC retention time: 17.3 min (500 nm)

ESI-MS = 73676 (MW calculated = 73673 g/mol)

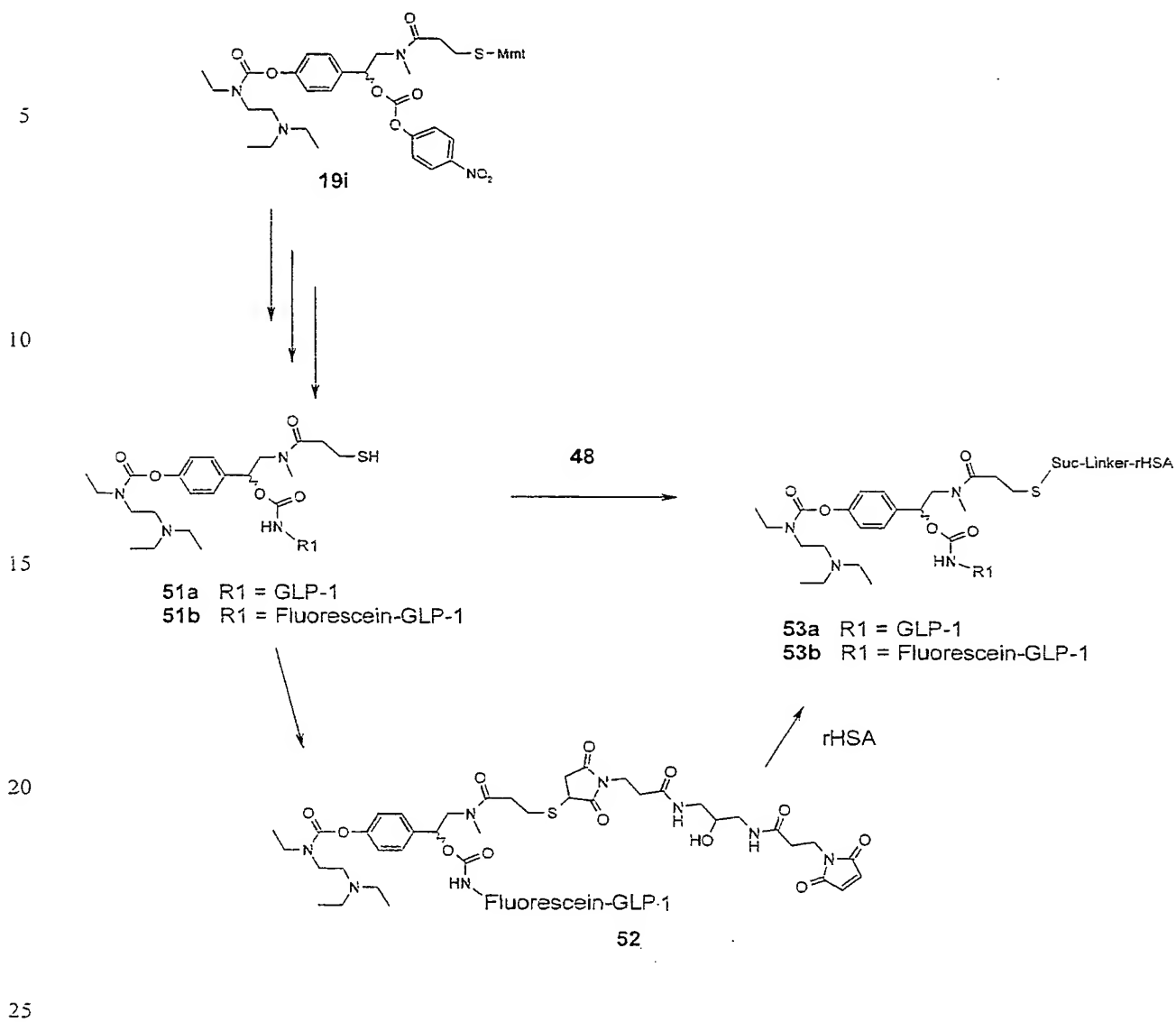
b)

20 SEC eluate of rHSA-maleimide 48 (241 μ l, 77.5 μ M, 18.7 nmol) was mixed with 20
 μ l 0.5 M sodium borate buffer pH 5.8. 14 μ l 1.41 mM 8f (19.6 nmol) in 1/1 (v/v)
acetonitrile/water were added and the mixture was incubated at RT for 10 min. 1.2 μ l
48.5 mM 3-maleimido propionic acid (58 nmol) in 1/1 (v/v) acetonitrile/water were
added and compound 50 was purified by SEC (column: Superdex 200, flow rate: 0.75
25 ml/min) using 10 mM HEPES buffer pH 7.4, 150 mM NaCl, 3 mM EDTA, and
0.005% Tween as mobile phase.

SEC retention time: 17.1 min (500 nm)

ESI-MS = 73698 (MW calculated = 73673 g/mol)

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Synthesis scheme of rHSA-linker-GLP-1 53a and 53bSynthesis of 51a

GLP(7-36) (sequence: HAEGTFTSDVSSYLEGQAAKEFLAWLVKGR-amide) was synthesized on Rink-amide resin employing fmoc-strategy (Specialty Peptide Laboratories, Heidelberg, Germany). N-terminal fmoc-protecting group was removed and the resin was washed with DCM and dried. 118 mg resin (0.11 mmol/g, 13.2 μmol) was suspended in a solution of 50 mg **19i** (53 μmol) in 750 μl dry DMSO and 22.4 μl DIEA. 2.1 μl pyridine was added and the mixture was shaken for 48 h at RT. After washing the resin 6 times each with DMF and DCM, cleavage of the peptide

from resin and removal of protecting groups was achieved with 96/2/2 (v/v/v) TFA/triethylsilane/water. Volatiles were removed under nitrogen flow and **51a** was purified by RP-HPLC and lyophilized.

5 **51a**: Yield 4.6 mg (9%)

MS: $[M+3H]^{3+} = 1251.0$ (MW calculated = 3750.3 g/mol)

Synthesis of **51b**

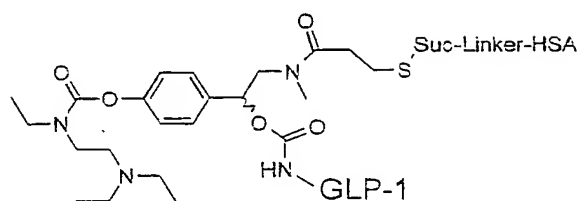
Lys28 ivDde side chain protected GLP(7-36) (sequence:
10 HAEGTFTSDVSSYLEGQAAKEFLAWLVK(ivDde)GR-amide) was synthesized on Rink-amide resin employing fmoc-strategy (Specialty Peptide Laboratories, Heidelberg, Germany). N-terminal fmoc-protecting group was removed and the resin was washed with DCM and dried. 50 mg resin (0.11 mmol/g, 5.5 μ mol) was suspended in a solution of 25 mg **19i** (26 μ mol) in 400 μ l dry DMSO and 11.2 μ l
15 DIEA. 1.1 μ l pyridine was added and the mixture was shaken for 48 h at RT. After washing the resin six times with DMF the ivDde protecting group was cleaved by incubating the resin 3 times with 5% hydrazine in DMF for 20 min. Fmoc-8-amino, 3,6-dioxaoctanoic acid was coupled according to the standard coupling cycle. Fmoc protecting group was removed and carboxy-fluorescein was coupled by incubating the
20 resin with 8 mg 5-(and-6)-carboxyfluorescein succinimidyl ester and 2 μ l DIEA for 60 min. Resin was washed six times each with DMF and DCM. Cleavage of the peptide from resin and removal of protecting groups was achieved with 96/2/2 (v/v/v) TFA/triethylsilane/water. Volatiles were removed under nitrogen flow. **51b** was used for the synthesis of **52** without further purification.
25 MS: $[M+3H]^{4+} = 1064.3$, $[M+2H]^{3+} = 1418.3$ (MW calculated = 4254 g/mol)

Synthesis of **52**

Raw material **51b** was dissolved in 500 μ l 1/1 (v/v) acetonitrile / 0.25 M sodium phosphate pH 7 and 8 mg N,N'-bis(3-maleimidopropionyl)-2-hydroxy-1,3-
30 propanediamine were added. The solution was stirred at RT for 15 min and **52** was purified by RP-HPLC and lyophilized.

52: Yield: 5.1 mg

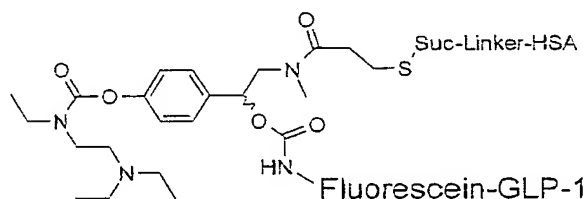
MS $[M+3H]^{4+} = 1162.8$, $[M+2H]^{3+} = 1549.4$ (MW calculated = 4645 g/mol)

Synthesis of compound 53a**53a**

- 5 30 μ l 1.57 mM **48** (47 nmol) in 10 mM HEPES buffer pH 7.4, 150 mM NaCl, 3 mM EDTA, and 0.005% Tween were mixed with 10 μ l 0.5 M sodium phosphate buffer pH 7.4. A mixture of 2 μ l DMSO and 12 μ l of 6.06 mM **51a** (73 nmol) in water/acetonitrile 9/1 (v/v) was added and the solution was incubated at RT for 30 min. **53a** was purified by SEC (column: Superdex 200, flow rate: 0.75 ml/min) using
- 10 10 mM phosphate buffer pH 7.4, 150 mM NaCl, and 0.005% Tween as mobile phase.

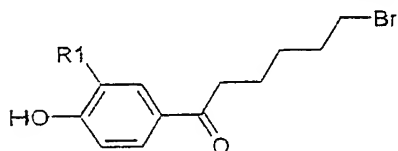
SEC retention time: 17.7 min (280 nm)

ESI-MS = 70745 (MW calculated = 70734 g/mol)

15 **Synthesis of 53b****53b**

- 100 μ l 3 mM **52** (300 nmol) in 9/1 50 mM sodium phosphate pH 7.0 / acetonitrile were mixed with 100 μ l 3 mM HSA (300 nmol) and the solution was incubated at RT
- 20 for 30 min. **53b** was purified by SEC (column: Superdex 200, flow rate: 0.75 ml/min) using 10 mM HEPES buffer pH 7.4, 150 mM NaCl, 3 mM EDTA, and 0.005% Tween as mobile phase.

SEC retention time: 17.7 min (500 nm)

Synthesis of compounds 54a and 54b

54a R1 = Me

5 **54b** R1 = H

AlCl₃ (1.05 eq) was suspended in DCM and 6-bromohexanoic acid chloride (1 eq) was added. After stirring at RT for 20 min o-cresol (1 eq) was added and the mixture was reacted at RT for 25 min. The reaction mixture was poured into ice water and
 10 extracted with ethyl acetate. The separated organic phase was dried over Na₂SO₄ and concentrated *in vacuo*. Product **54a** was purified by silica gel column chromatography using heptane/ethyl acetate (4/1) as mobile phase.

54b was synthesized as described above using 6-bromohexanoic acid chloride and phenol.

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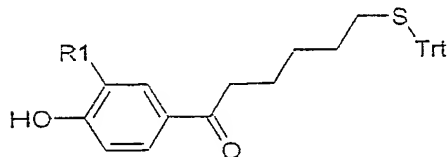
54a: Yield 3.7 g (33%)

MS [M+H]⁺ = 285.1 and 287.2 (MW+H calculated = 386.2 g/mol)

54b: Yield 620 mg (15%)

MS [M+H]⁺ = 271.2 (MW calculated = 271.0 g/mol)

20

Synthesis of compounds 55a and 55b

25 **55a** R1 = Me

55b R1 = H

DBU (105 μ l, 701 μ mol) was added to a solution of bromide **54a** (105 mg, 369 μ mol) and tritylthiol (204 mg, 738 μ mol) in 50 ml dry DMSO. The reaction mixture was stirred at RT for 40 min and acidified with 1 N H_2SO_4 . The aqueous layer was extracted with ethyl acetate and evaporated. **55a** was purified by RP-HPLC.

55b was synthesized according to the same protocol using **54b** (180 mg, 0.66 mmol).

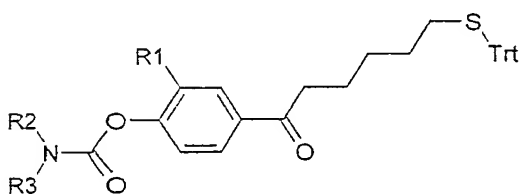
55a: Yield 173 mg (97%)

MS $[\text{M}+\text{Na}]^+ = 503.6$ (MW+Na calculated = 503.7 g/mol)

55b: Yield 160 mg (85%)

MS $[\text{M}+\text{Na}]^+ = 489.5$ (MW+Na calculated = 489.3 g/mol)

Synthesis of compounds **56a** and **56b**



56a R1 = Me, R2 = R3 = 3-(dimethylamino)propyl,

56b R1 = H, R2 = Me, R3 = 3-(N-ethyl-N-methylamino)propyl

56a was prepared from **55a** (9 mg, 19 μ mol), p-nitrophenyl-chloroformate and bis(3-dimethylamino-propyl)amine (21 μ l, 94 μ mol) as described for compound **16a**.

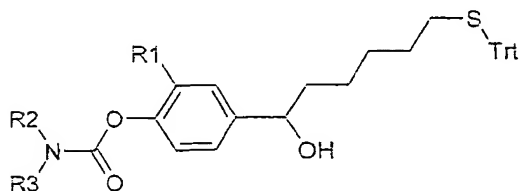
56b was synthesized from **55b** (160 mg, 0.34 mmol), p-nitrophenyl-chloroformate and N-ethyl-N,N'-dimethyl-1,3-propanediamine (**15c**) as described for compound **16a**.

56a: Yield 12 mg (70%) as TFA salt

MS $[\text{M}+\text{Na}]^+ = 716.8$ (MW+Na calculated = 717.0 g/mol)

56b: Yield 80 mg (32%) as TFA salt

MS $[\text{M}+\text{Na}]^+ = 645.6$ (MW+Na calculated = 645.4 g/mol)

Synthesis of compounds 57a and 57b

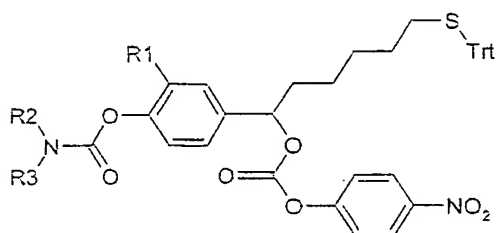
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57a R1 = Me, R2 = R3 = 3-dimethylamino-propyl,**57b** R1 = H, R2 = Me, R3 = 3-(N-ethyl-N-methylamino)propyl

57a and **57b** were synthesized from **56a** (12 mg, 13 μ mol, double TFA salt) and **56b**
 10 (80 mg, 110 μ mol, TFA salt), respectively, as described for compound **16g**.

57a: Yield 9 mg (75%) as TFA saltMS $[M+Na]^+ = 719.0$ (MW+Na calculated = 718.7 g/mol)**57b:** Yield 60 mg (75%) as TFA salt

15 MS $[M+Na]^+ = 647.4$ (MW+Na calculated = 647.4 g/mol)

Synthesis of compounds 58a and 58b

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58a R1 = Me, R2 = R3 = 3-(dimethylamino)propyl,**58b** R1 = H, R2 = Me, R3 = 3-(N-ethyl-N-methylamino)propyl

25 **57a** (1 eq, 8mg, 9 μ mol), 4-nitrophenyl chloroformate (3.5 eq, 6 mg, 30 μ mol), DIEA
 (6 eq, 9 μ l, 52 μ mol), and DMAP (1 eq, 1 mg, 9 μ mol) were stirred in 1 ml dry DCM

at RT for 45 min under nitrogen atmosphere. The volatiles were evaporated and acetic acid was added. The mixture was dissolved in 1/1 (v/v) acetonitrile/water and the carbonate **58a** was purified by RP-HPLC.

Carbonate **58b** was prepared likewise from **57b** (135 mg, 0.18 mmol).

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58a: Yield 7 mg (70%) as TFA salt

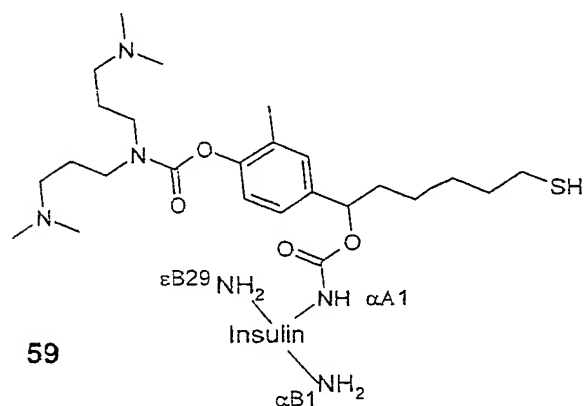
MS $[M+Na]^+ = 883.8$ (MW+Na calculated = 884.1 g/mol)

58b: Yield 110 mg (77%) as TFA salt

MS $[M+Na]^+ = 812.4$ (MW+Na calculated = 812.5 g/mol)

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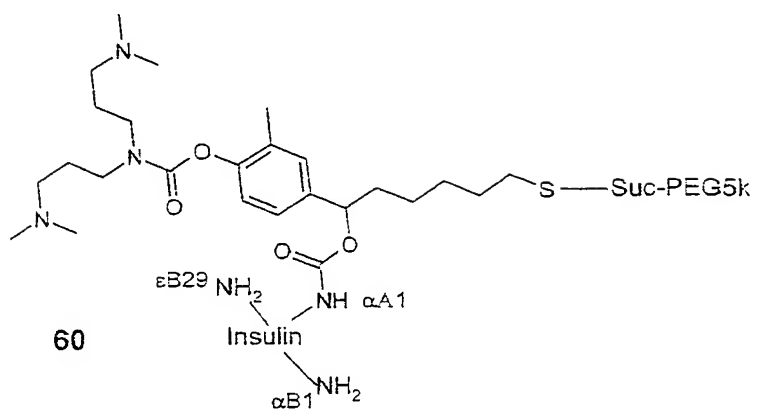
Synthesis of compound **59**



15 Rh-Insulin (44.5 mg, 7.7 μ mol), carbonate **58a** (1 eq, 7 mg, 6.4 mmol), DIEA (15 μ l, 88 μ mol) and DMAP (1.5 mg, 12 μ mol) in 0.3 ml DMSO were reacted at RT for 30 min. The reaction mixture was neutralized with acetic acid and diluted with 1/1 (v/v) acetonitrile/water. RP-HPLC purification gave the appropriate Trt-protected intermediate.

20 After lyophilization, the Trt-protected intermediate was mixed with 95/5 (v/v) TFA/triethylsilane and stirred for 5 min. Volatiles were removed under nitrogen flow and **59** was purified by RP-HPLC and lyophilized. Position of insulin modification was verified by DTT reduction and MS analysis.

25 **59**: MS $[M+3H]^{3+} = 2095.5$ $[M+4H]^{4+} = 1572.2$ (MW calculated = 6288 g/mol)

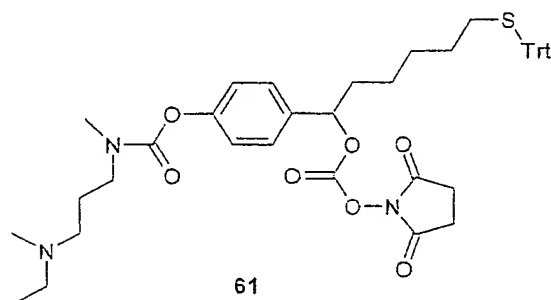
Synthesis of compound 60

5 60 was prepared from 59 (0.17 μmol) as described for compound 9a.

60: SEC retention time: 19.5 min

Synthesis of compound 61

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57b (70 mg, 90 μmol), DSC (161 mg, 630 μmol), DIEA (192 μl , 1.1 mmol), and DMAP (11 mg, 90 μmol) were stirred in 1 ml dry acetonitril at RT for 14 h under nitrogen atmosphere. The volatiles were evaporated and acetic acid was added. The mixture was dissolved in 1/1 (v/v) acetonitrile/water and the carbonate 61 was purified by RP-HPLC.

61: Yield 40 mg (51%) as TFA salt

20 MS $[M+\text{Na}]^+ = 788.4$ (MW+Na calculated = 788.5 g/mol)

CCN(C)CCNC(=O)Oc1ccc(cc1)C(OC(=O)O=C2CCCC2=O)CCCCSSc3ccnc(c3)[N+](=O)[O-]

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$$\text{MS } [\text{M}+\text{Na}]^+ = 700.9 \text{ (MW}+\text{Na calculated} = 701.4 \text{ g/mol)}$$

63

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weighed compounds were replaced by water and subsequently by acetate buffer (25 mM, pH 4.2, 0.005% Tween 20) by ultrafiltration using Centricon 5 filter (cutoff 5 kDa). 8 μ l (80 nmol) 10 mM DTT in 25 mM acetate buffer pH 4.2, 0.005% Tween was added and incubated at RT for 30 min. Low molecular weight compounds were removed by ultrafiltration using Centricon 5 filter and 25 mM acetate buffer pH 4.2, 0.005% Tween as eluate. After concentration to a volume of 100 μ l (Centricon 5) 20 μ l (100 nmol) 5 mM maleimide-PEG5k in water and 80 μ l 0.5 M phosphate buffer pH 7.0 were added. The mixture was incubated at RT for 5 min. Monoconjugate **63** was separated by SEC (column: Superdex 200, flow rate: 0.75 ml/min) using 10 mM phosphate buffer pH 7.4, 150 mM NaCl, and 0.005% Tween 20 as mobile phase. The collected eluate (approximately 1.0 ml) was diluted with 0.5 ml buffer containing 0.05% NaN₃ and directly used for release rate determination.

63: SEC retention time: 17.5 min

Release of insulin or fluorescein-insulin from conjugates in buffer pH 7.4

Release of (fluorescein)-insulin from (fluorescein)-insulin conjugates **9a** to **9h**, **21a** to **21f**, **30a**, **30b**, **43**, **50**, and **60**, release of fluorescein-GLP-1 from **53b**, and release of rhGH from **63** was effected by linker hydrolysis in aqueous buffer pH 7.4. Collected SEC eluates of (fluorescein)-insulin conjugates (see above), fluorescein-GLP-1 conjugate and rhGH-conjugate, respectively, were incubated at 37 °C and samples were taken at time intervals and analyzed by RP-HPLC (insulin conjugates) or SEC (rhGH conjugate, fluorescein insulin conjugates and fluorescein-GLP-1 conjugate) and UV detection at 215 or 280 nm or VIS detection at 500 nm. Peaks correlating with the retention time of native insulin, fluorescein-insulin, fluorescein-GLP-1, and rhGH, respectively, were integrated and plotted against incubation time, and curve-fitting software was applied to estimate the corresponding halftime of release.

Release of insulin from hydrogel conjugates **45** and **46**

4 mg of **45** or 2 mg **46** were weighed into a test tube and incubated with 1 ml 10 mM HEPES buffer pH 7.4, 150 mM NaCl, 0.005% Tween at 37°C. 45 μ l samples were

taken at different time intervals and quantitatively analyzed for rh-insulin by a RP-HPLC assay. The rh-insulin peaks were integrated and rh-insulin concentration was obtained from a standard curve. A first order release kinetic was fitted to the data points to give the linker half life.

5

MS-analysis of released insulin from compound 9a, 9b and 30a

Samples of buffer released insulin (see above) were analyzed by mass spectrometry. Figure 11 shows the mass spectra of released insulin from compound 9a, 9b, and 30a. The mass spectrum of insulin released from compound 9a clearly shows a major side product (indicated by arrows), corresponding to the irreversibly pentanoyl-modified insulin. In this case, removal of the pentanoyl masking group was not by hydrolysis but by acyl transfer to the insulin. The mass spectrum of insulin released from compound 9b and 30a shows no modification.

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Release of fluorescein-insulin from conjugate 9d and 9e in 80% human plasma

Release of fluorescein-insulin from 9d or 9e was effected by hydrolysis in 80% human plasma in 20 mM HEPES pH 7.4 at 37 °C. Samples were taken at time intervals and analyzed by SEC and VIS detection at 500 nm. Peaks correlating with the retention time of fluorescein-insulin were integrated and plotted against incubation time, and curve-fitting software was applied to estimate the corresponding halftime of release.

25 Table: Polymeric prodrug hydrolysis

compound	t _{1/2} buffer pH 7.4	t _{1/2} human plasma
9a	40 h	nd
9b	55 h	nd
9c	4.5 d	nd
9d	7 h	4 h
9e	55 h	30 h
9f	90 h	nd
9g	37 h	nd

9h	88 h	nd
21a	64 d	nd
21b	8 d	nd
21c	52 d	nd
21d	29 d	nd
21e	100 d	nd
21f	83 h	nd
30a	17 d	nd
30b	> 70 d	nd
43	4 h	nd
45	7 d	nd
46	4 d	nd
50	57 h	nd
53b	19 h	nd
60	10 d	nd
63	51 d	nd

nd = not determined

The foregoing is considered illustrative of the principles of the invention and since numerous modifications will occur to those skilled in the art, it is not intended to limit
5 the invention to the exact construction and operation described. All suitable modifications and equivalents fall within the scope of the claims.

Abbreviations:

	Boc	t-butyloxy carbonyl
	DBU	1,3-diazabicyclo[5.4.0]undecene
	DCM	dichloromethane
5	(iv)Dde	1-(4,4-dimethyl-2,6-dioxo-cyclohexyliden)3-methyl-butyl
	DIC	diisopropylcarbodiimide
	DIEA	diisopropylethylamine
	DMAP	dimethylamino-pyridine
	DMF	N,N-dimethylformamide
10	DMSO	dimethylsulfoxide
	Dpr	diaminopropionic acid
	DSC	disuccinidylcarbonate
	EDTA	ethylenediaminetetraacetic acid
	Et	ethyl
15	eq	stoichiometric equivalent
	fmoc	9-fluorenylmethoxycarbonyl
	Fmoc-Ado-OH	Fmoc-8-amino-3,6-dioxaoctanoic acid
	HFIP	hexafluoroisopropanol
	HEPES	N-(2-hydroxyethyl) piperazine-N'-(2-ethanesulfonic acid)
20	HOBt	N-hydroxybenzotriazole
	LCMS	mass spectrometry-coupled liquid chromatography
	Mal	maleimido propionyl
	Me	methyl
	Mmt	4-methoxytrityl
25	MS	mass spectrum
	MW	molecular mass
	Npys	3-nitro-2-pyridinesulfonyl
	PyBOP	benzotriazole-1-yl-oxy-tris-pyrrolidino-phosphonium hexafluorophosphate
30	rHSA	recombinant human serum albumin
	rhGH	recombinant human growth hormone
	RP-HPLC	reversed-phase high performance liquid chromatography
	RT	room temperature

	SEC	size exclusion chromatography
	Suc	succinimidopropionyl
	TES	triethylsilane
	TFA	trifluoroacetic acid
5	THF	tetrahydrofuran
	UV	ultraviolet
	VIS	visual
	Z	benzyloxycarbonyl

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